

Comparison and Classification of Protein Structures

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Protein Data Bank (PDB)

Worldwide Protein Data Bank (wwPDB)

The screenshot shows the wwPDB homepage with a green and white color scheme. The header includes the PDB logo and navigation links. A large central banner celebrates the 100,000th structure deposited, featuring a digital display showing '100147'. The text on the banner states: 'The Worldwide Protein Data Bank (wwPDB) consists of organizations that act as deposition, data processing and distribution centers for PDB data. Members are: RCSB PDB (USA), PDBe (Europe) and PDBj (Japan). The wwPDB's mission is to maintain a single PDB archive of macromolecular structural data that is freely and publicly available to the global community.' Below the banner, there are sections for 'Access the PDB FTP', 'New Deposition and Annotation System', 'Validation Reports', 'Deposit Data to the PDB', 'Search for Structures', 'PDB Archive Snapshots', 'Instructions to Journals', 'Workshops and Task Forces', 'Past Symposia', and 'Full News'. A sidebar on the left contains links to various PDB resources.

Protein Data Bank Japan (PDBj)

The screenshot shows the PDBj homepage with a blue and white color scheme. The header includes the PDBj logo and navigation links. A large central banner displays the number '100693' and the text 'PDBj Protein Data Bank Japan'. Below the banner, there are sections for 'ホーム' (Home), '初めての利用者のためのガイド' (Guide for first-time users), 'サービスを探す' (Find services), '最新情報' (Latest information), and 'サービス&ソフトウェア' (Services & Software). The 'サービスを探す' section lists various services such as PDB, BMRB, EMDB, and others. The '最新情報' section contains news items about the 100,000th structure and other PDB milestones. The 'サービス&ソフトウェア' section lists various software tools and services available for PDB data.

The primary database of biological macromolecular structures

An example of PDB entries

100693
PDBj
Protein Data Bank Japan

English 日本語 繁体中文 繁體中文 한국어

1GOF

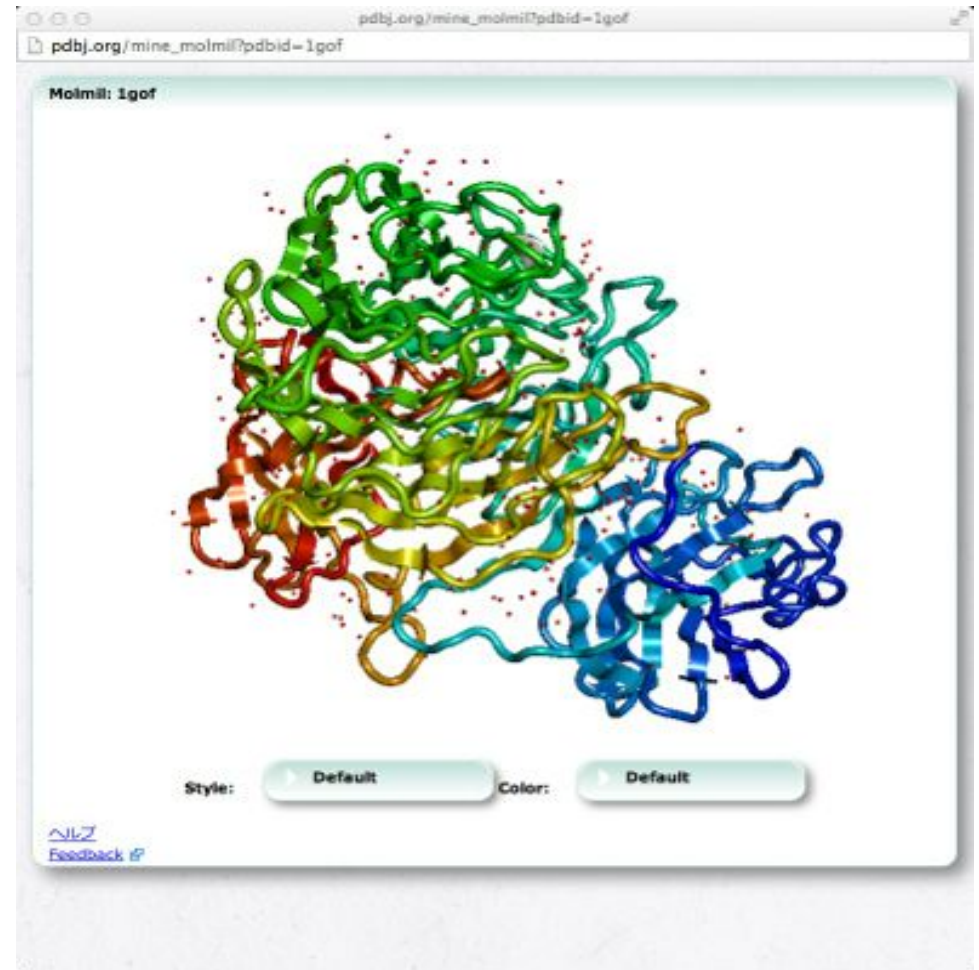
NOVEL THIOETHER BOND REVEALED BY A 1.7 ANGSTROMS CRYSTAL STRUCTURE OF GALACTOSE OXIDASE

1GOF の概要

分子名称	GALACTOSE OXIDASE (E.C.1.1.3.9) (PH 4.5)
機能のキーワード	OXIDOREDUCTASE(OXYGEN(A))
由来する生物種	Hypomyces rosellus
ポリマー鎖の合計数	1
分子量の合計	68765.00
著者	Ito, N., Phillips, S.E.V., Knowles, P.F. (登録日: 1993-09-30, 公開日: 1994-01-31, 最終更新日: 2011-07-13)
引用文献	Ito, N., Phillips, S.E., Stevens, C., Ogel, Z.B., Mulherson, M.J., Keen, J.N., Yadan, K.D., Knowles, P.F. Novel thioether bond revealed by a 1.7 Å crystal structure of galactose oxidase. Nature, 359: 87-90, 1991 PubMed: 2002900 DOI: 10.1038/359087a0 MolBiochemBiophys 1991 MolBiochemBiophys 1991
実験手法	X-RAY DIFFRACTION (1.7 Å)

他の表示画像 (非対称単位)

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Protein Structure Comparison

```
[BEGIN ALIGNMENT]
      : E1          H1          E2          H2          E3
SecA : EEEEE S  HHHHHHHHHHHHS  SSEEEEE GGGHHHHHHH  TTEEEE
3 :RTFFVGGNFKLNGSKQSIKEIVERLNTASIPENVEVVICPPATYLDYSVSLVKKPQVTVG: 62
   * * * * * * * * * * * * * * * * * * * * * * * * * * *
4 :RKFFVGGNWKMNMGDKKSLGELIHTLNGAKLSADTEVVCGAPSIYLDFAEQKL-DAKIGVA: 62
SecB : EEEEE S  HHHHHHHHHHHHS  TTEEEEE GGGHHHHHHHS- TTSEEE
      : E1          H1          E2          H2          -   E3

      :          H3          E4          H4          H5          E
SecA :ES  SSSSSS TT  HHHHHHTT  EEEE  HHHHHHS  HHHHHHHHHHHHTT E
63 :AQNAYLKASGAFTGENSVDIKDVGAQWVILGHSERRSYFHEDDKFIADKTKFALGQGVG: 122
   * * * * * * * * * * * * * * * * * * * * * * * * * * *
63 :AQNCYKVPKGAFTGEISPAMIKDIGAAWVILGNPERRHVFGESDELIGQKVAHALAEGLG: 122
SecB :ES  SSSSSS TT  HHHHHHTT  EEEE  HHHHHTS  HHHHHHHHHHHHTT E
      :          H3          E4          H4          H5          E

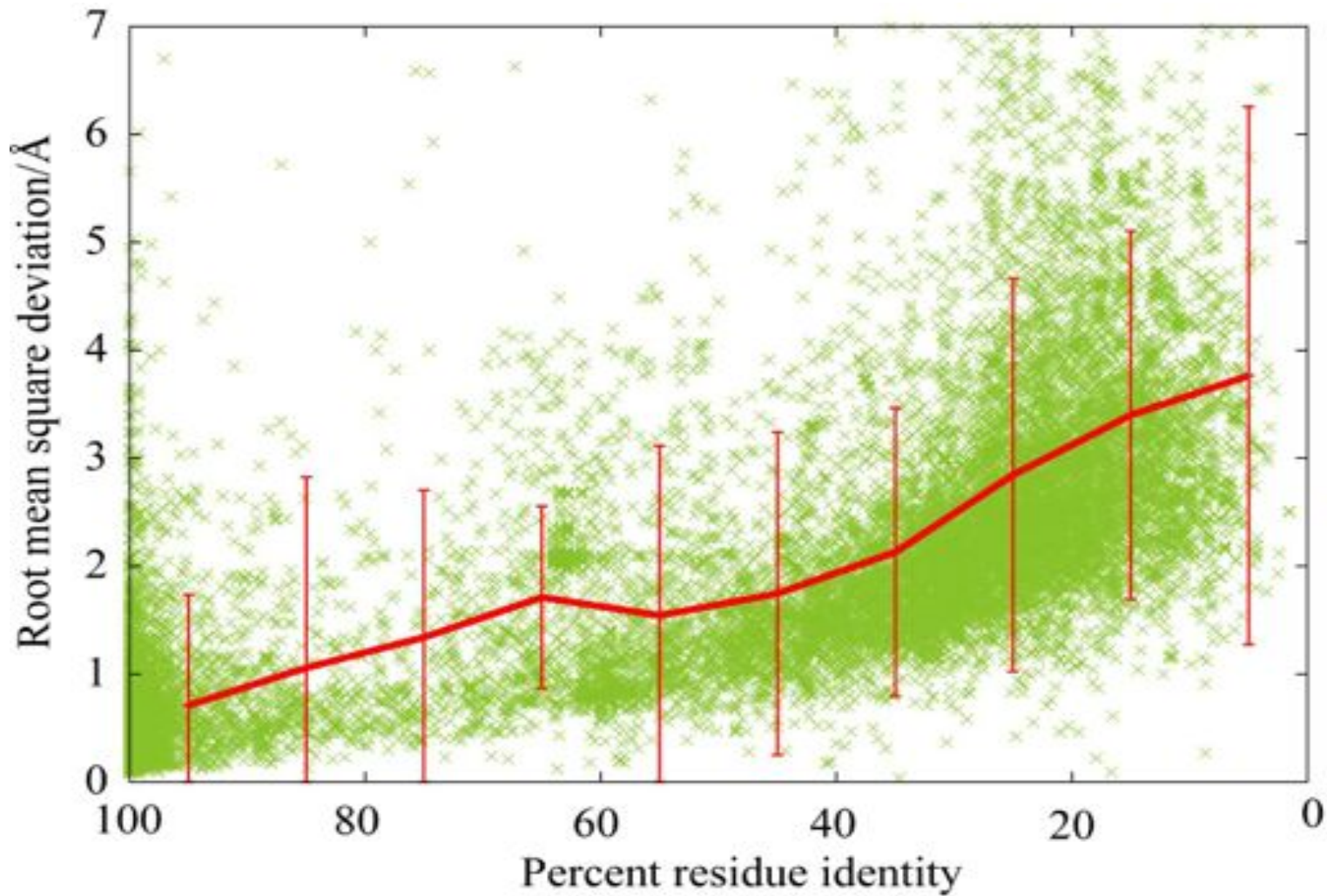
      :5          H6          H7          E6          H8
SecA :EEEE  HHHHHHTT  HHHHHHHHHHHHHH S  TTEEEEE GGGTTTS  HHHHH
123 :VILCIGETLEEKAGKTLDVVERQLNAVLEEVDWNTNVVAYEPVWAIGTGLAATPEDAQ: 182
   ** * * * * * * * * * * * * * * * * * * * * * * * * * *
123 :VIACIGEKLDEREAGITEKVVFQTKAIADNVKDWKVVLAYEPVWAIGTGKTATPQQAQ: 182
SecB :EEEE  HHHHHHTT  HHHHHHHHHHHHHHTT S  GGGEEEE  GGGTTTS  HHHHH
:5          H6          H7          E6          H8

      :          H9          E7          E8          H10
SecA :HHHHHHHHHHHHHH HHHHHH  EEEESS  TTTGGGGTT TT  EEEESGGGGSTTHHH
183 :DIHASIRKFLASKLGDKAASELRILYGGSSANGSNAVTFKDKADVDGFLVGASLKPEFVD: 242
   * * * * * * * * * * * * * * * * * * * * * * * * * * *
183 :EVHEKLRGWLKTHVSDAVAQSTRIIYGGSVTGGNCKELASQHDVDGFLVGASLKPEFVD: 242
SecB :HHHHHHHHHHHHHT HHHHHH  EEEESS  TTTHHHHHTSTT  EEEESGGGGSTTHHH
      :          H9          E7          H10          E8          H11

      :
SecA :HHHTT
243 :IINSRN: 248
     ***
243 :IINAKH: 248
SecB :HHHTT
      :
```



Sequence & structure similarities

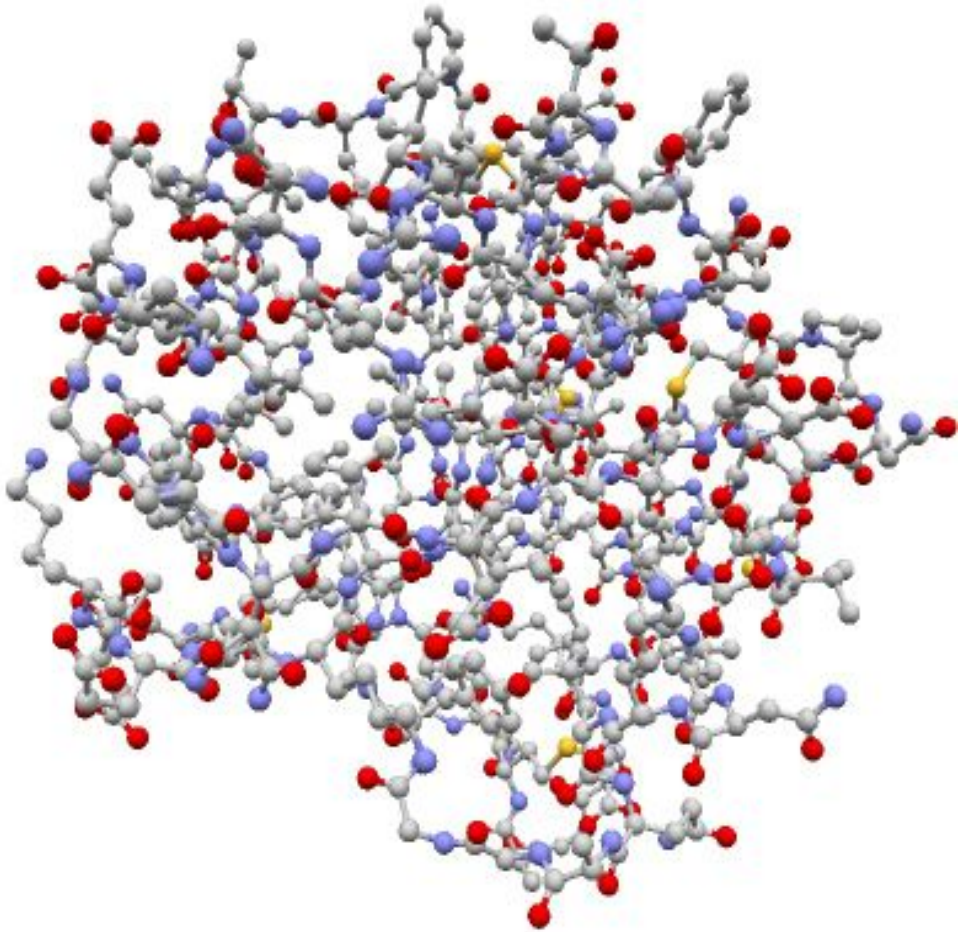


(『タンパク質の立体構造入門』図 4.1)

Methods of structure comparison

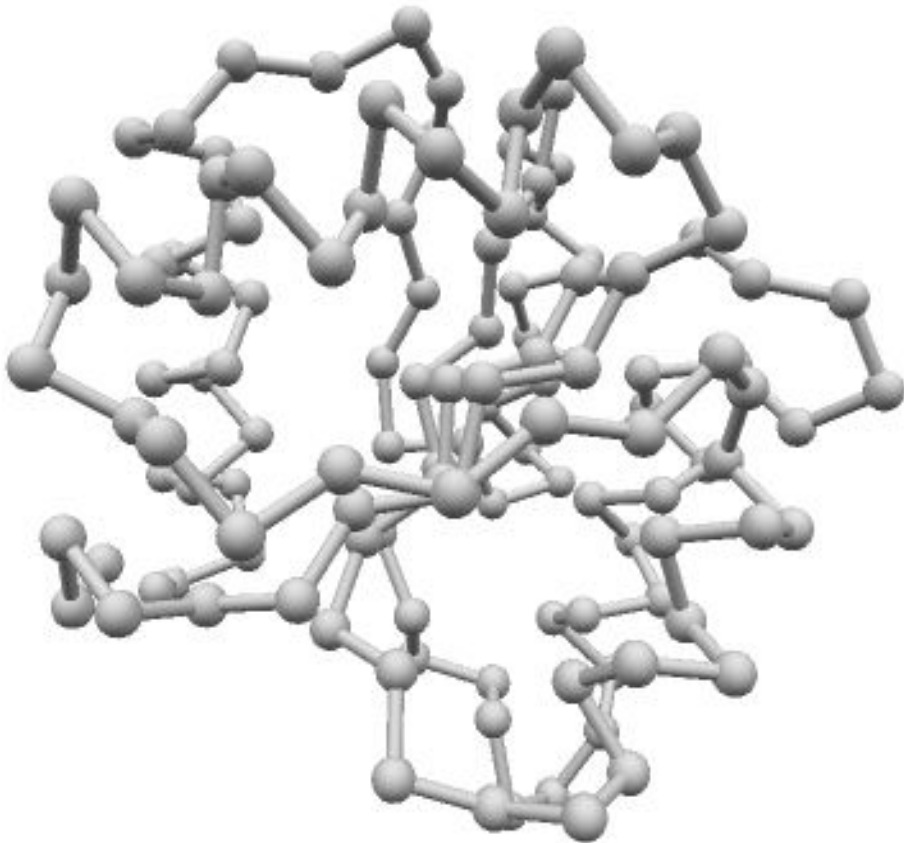
- Visual inspection(!)
 - The “best” method if you are well-trained.
- Algorithms
 - It's an NP-hard problem, so there are a number of approximate methods based on various representations:
 - secondary structure elements (SSE)
 - Amino acid residues
 - Atoms
 - Molecular surface

Representation: all atoms



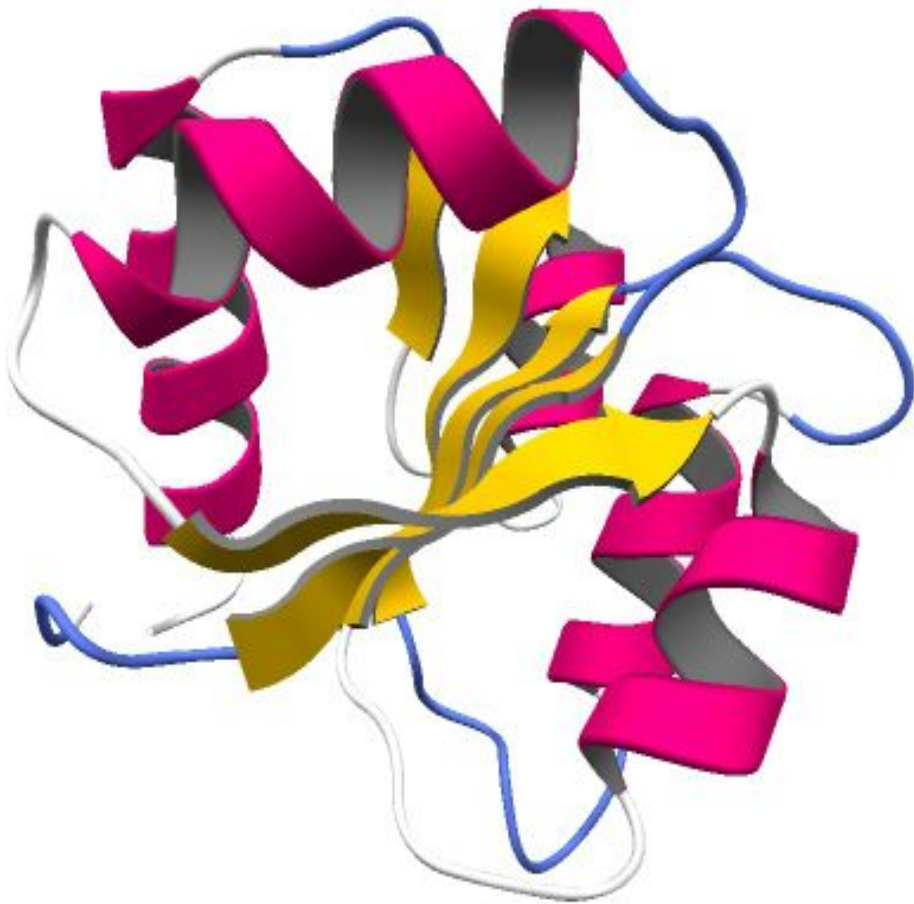
- Dealing with all atoms...
- is difficult. So usually only substructures are treated in this representation.

Representation: Backbone



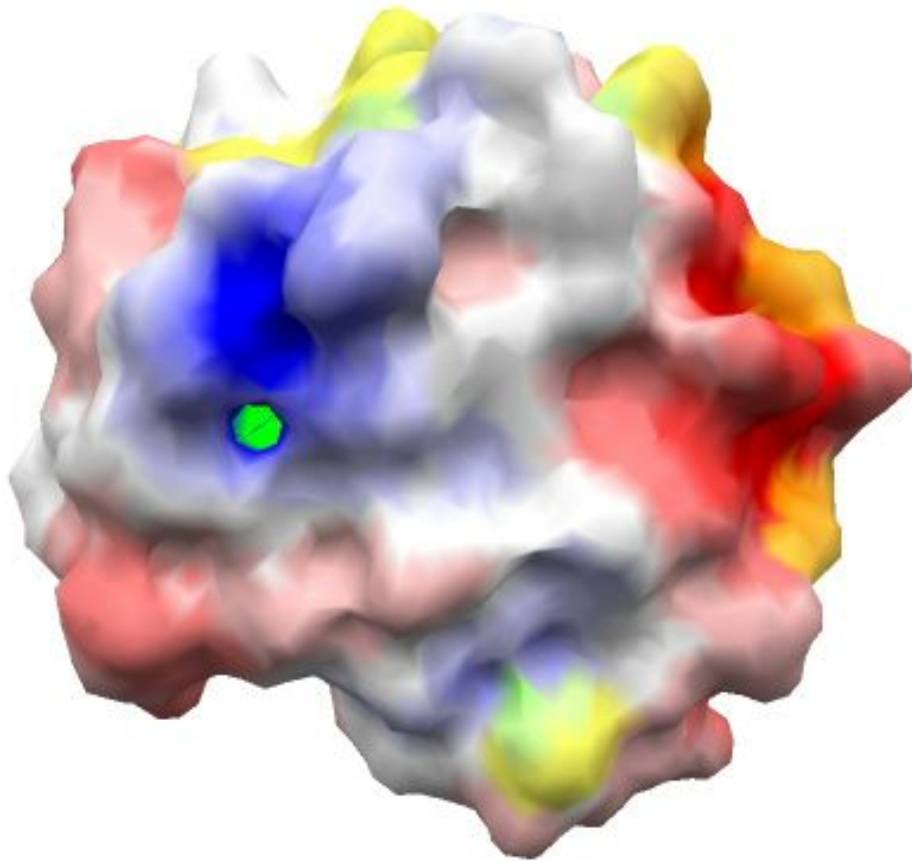
- Using only $C\alpha$ or $C\beta$ atoms to reduce computational costs.
- Also compatible with sequence alignment (1 atom / residue)
- Still computationally demanding.

Representation: 2ndary structures



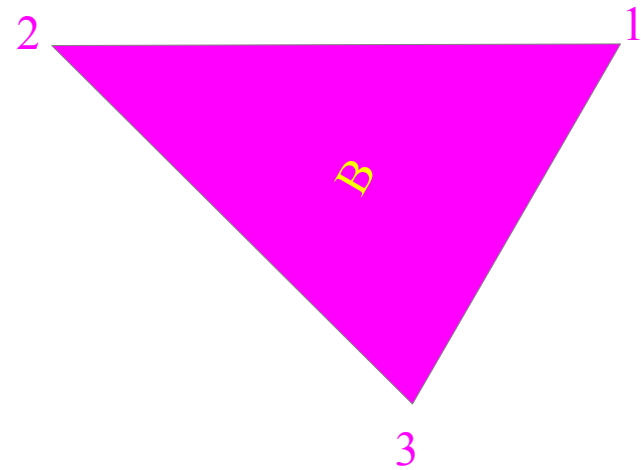
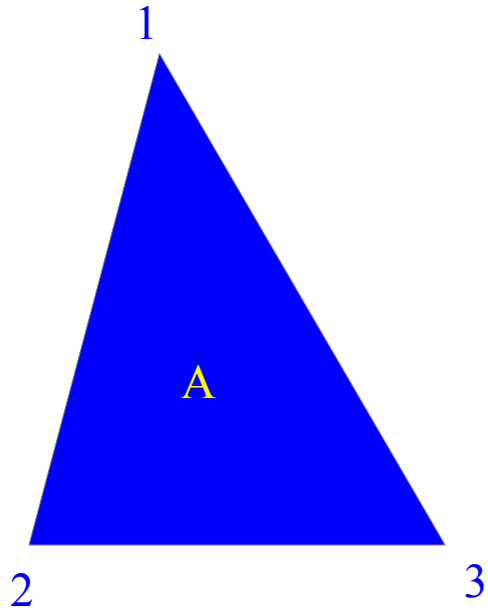
- α helices and β strands as vectors.
- Suitable for finding topological similarities.
- Less cost.

Representation: Molecular surface



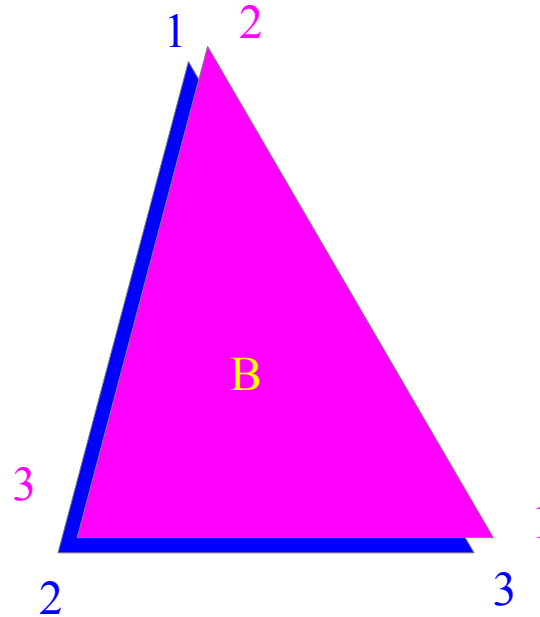
- Protein structure from the view point of a water molecule(?)
- Often used for mapping electrostatic potentials & hydrophathy on the structure.

Basic ideas



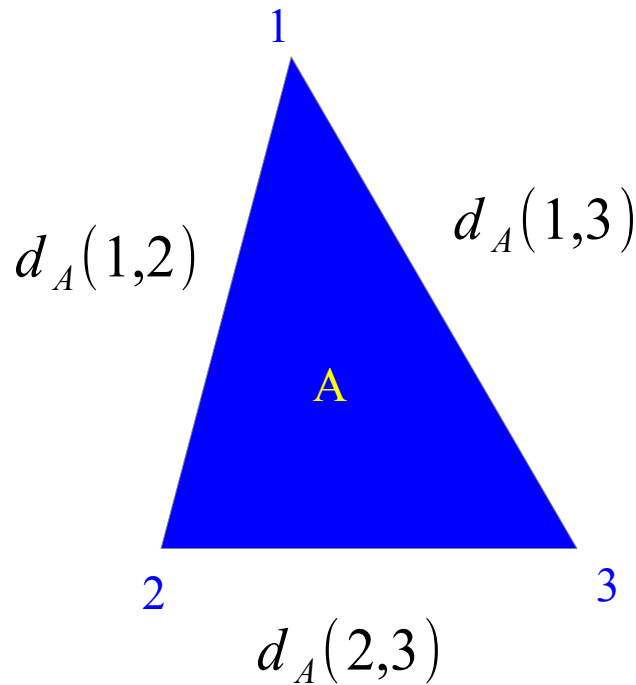
How do you tell the congruence of two triangles?
(Vertex numbers do not match!)

Method 1: Coordinate-based

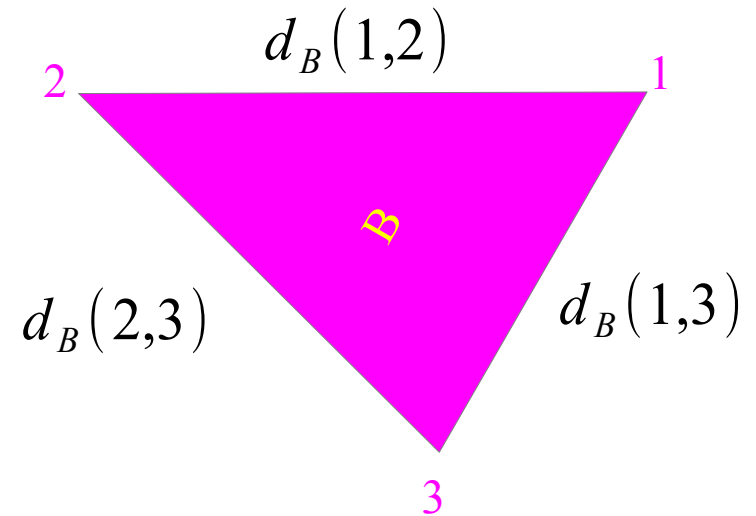


- Actually try to superimpose them!
- Infinite combinations of “translation” & “rotation”.

Method 2: Distance-based



	$d_A(1,2)$	$d_A(1,3)$
$d_A(2,1)$		$d_A(2,3)$
$d_A(3,1)$	$d_A(3,2)$	

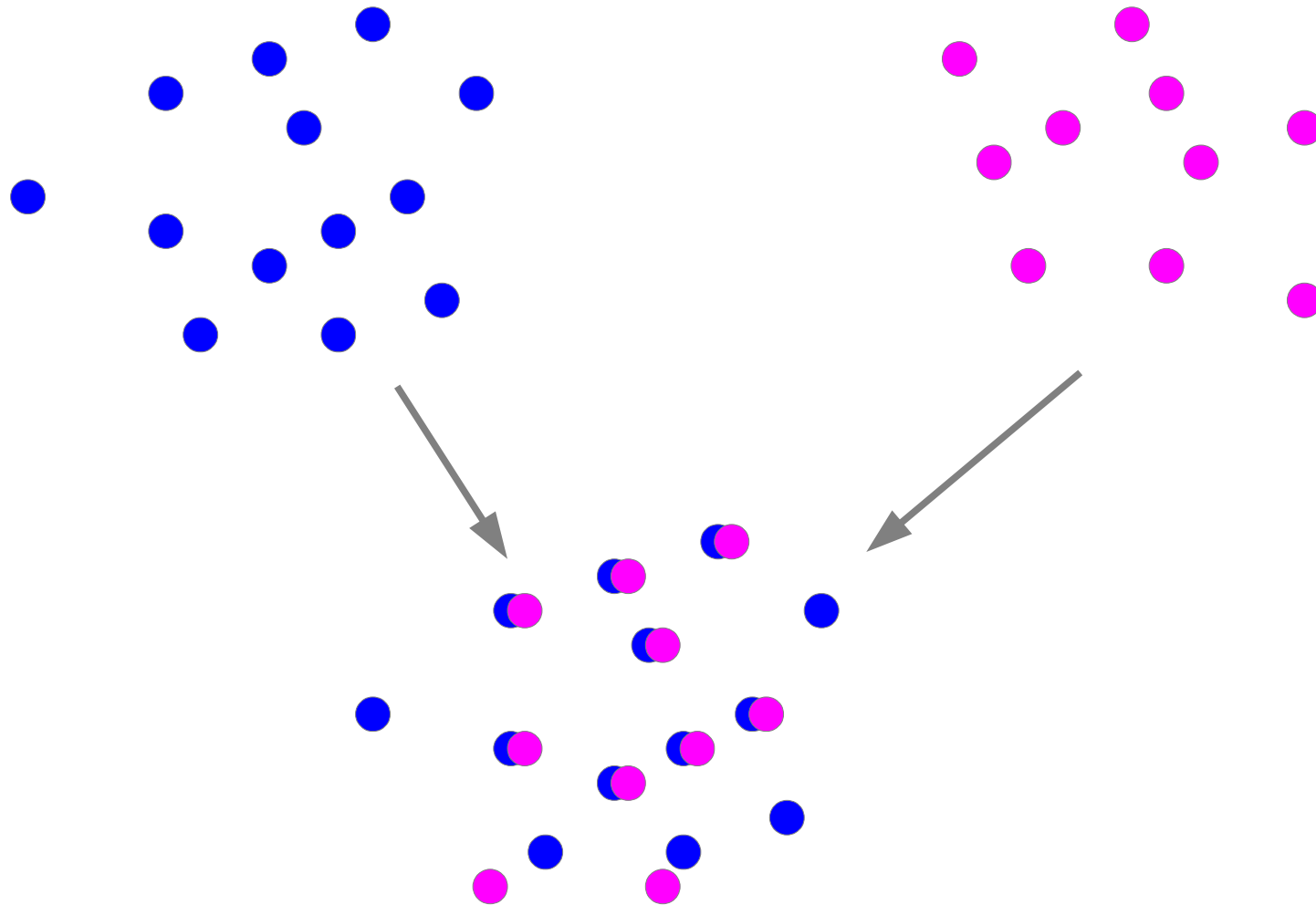


	$d_B(1,2)$	$d_B(1,3)$
$d_B(2,1)$		$d_B(2,3)$
$d_B(3,1)$	$d_B(3,2)$	

Find pairs $(i,j),(k,l)$ that satisfy $|d_A(i, j) - d_B(k, l)| = 0$

How many possibilities are there?

A little more complicated objects



Summary of comparison methods

- Translation & rotation
 - “Coordinate-based method”
 - Infinite possibilities.
- Comparing the distances between vertices
 - “Distance-based method”
 - Exponentially increasing possibilities.
- In any case, it's a tough problem!

Coordinate-based method, theory

Let A , B and C be metric spaces:

$$A = (x_1^A, x_2^A, \dots, x_M^A) \quad B = (x_1^B, x_2^B, \dots, x_N^B)$$

$$A \xrightarrow{f} C$$

$$B \xrightarrow{g} C$$

The points in A and B are transformed into C , so the distance between two points, one in A and the other in B can be measured in C .

$$D(A, B) = \sum_{(i,j)} d_C(f(x_i^A), g(x_j^B))$$

The Problem: Find the set of combinations of (i,j) that minimizes this distance.
But how do we define f and g ?

Best-fitting problem

Easy case first. Assume the alignment is already known.

$$A = (x_1^A, x_2^A, \dots, x_M^A) \quad B = (x_1^B, x_2^B, \dots, x_M^B) \quad (\text{the same number of points})$$

$$\text{For all } i=1, \dots, M, (x_i^A, x_i^B) \quad (\text{The } i\text{-th atom in A } \Leftrightarrow \text{The } i\text{-th atom in B})$$

$$\sum_{i=1}^M x_i^A = 0, \sum_{i=1}^M x_i^B = 0 \quad (\text{Both centers of mass are at the origin})$$

$$D(A, B) = \sqrt{\frac{1}{M} \sum_{i=1}^M |x_i^A - R x_i^B|^2} \quad (\text{Rotate B by the rotation matrix R})$$

Now the problem is finding the matrix R (least-square fitting).

This can be solved analytically (Euler angles, singular value decomposition, quaternions)

Coordinate-based method in practice

- Impossible to try infinite number of transformations
- 3 linearly independent points define a frame.
 - $N!/(N-3)! = N(N-1)(N-2)$
- Consider all combination from two structures
 - $M(M-1)(M-2) \times N(N-1)(N-2)$
 - $M=N=100 \Rightarrow 941,288,040,000$ combinations
- It's finite, but huge!

Coordinate frame based on 3 points

$$A = (x_1^A, x_2^A, \dots, x_M^A)$$

$$(x_i^A, x_j^A, x_k^A) \quad \text{3 points}$$

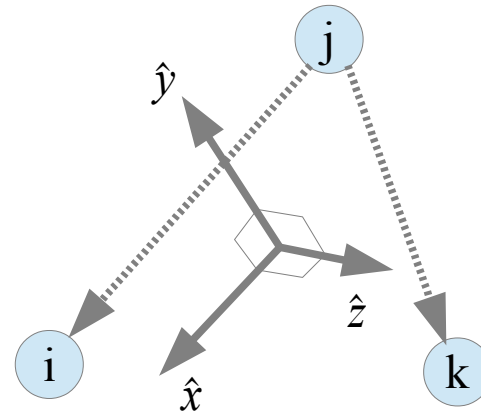
$$\hat{x} = \frac{1}{\|x_i^A - x_j^A\|} (x_i^A - x_j^A) \quad \text{X axis}$$

$$\hat{y} = \frac{1}{\|x_k^A - x_j^A\|} \hat{x} \times (x_k^A - x_j^A) \quad \text{Y axis}$$

$$\hat{z} = \hat{x} \times \hat{y} \quad \text{Z axis}$$

$$O = \frac{1}{3} (x_i^A + x_j^A + x_k^A) \quad \text{Origin}$$

$$\hat{x}_a^A = (\hat{x} \cdot (x_a^A - O), \hat{y} \cdot (x_a^A - O), \hat{z} \cdot (x_a^A - O)), a = 1, \dots, M \quad \text{Transformation}$$

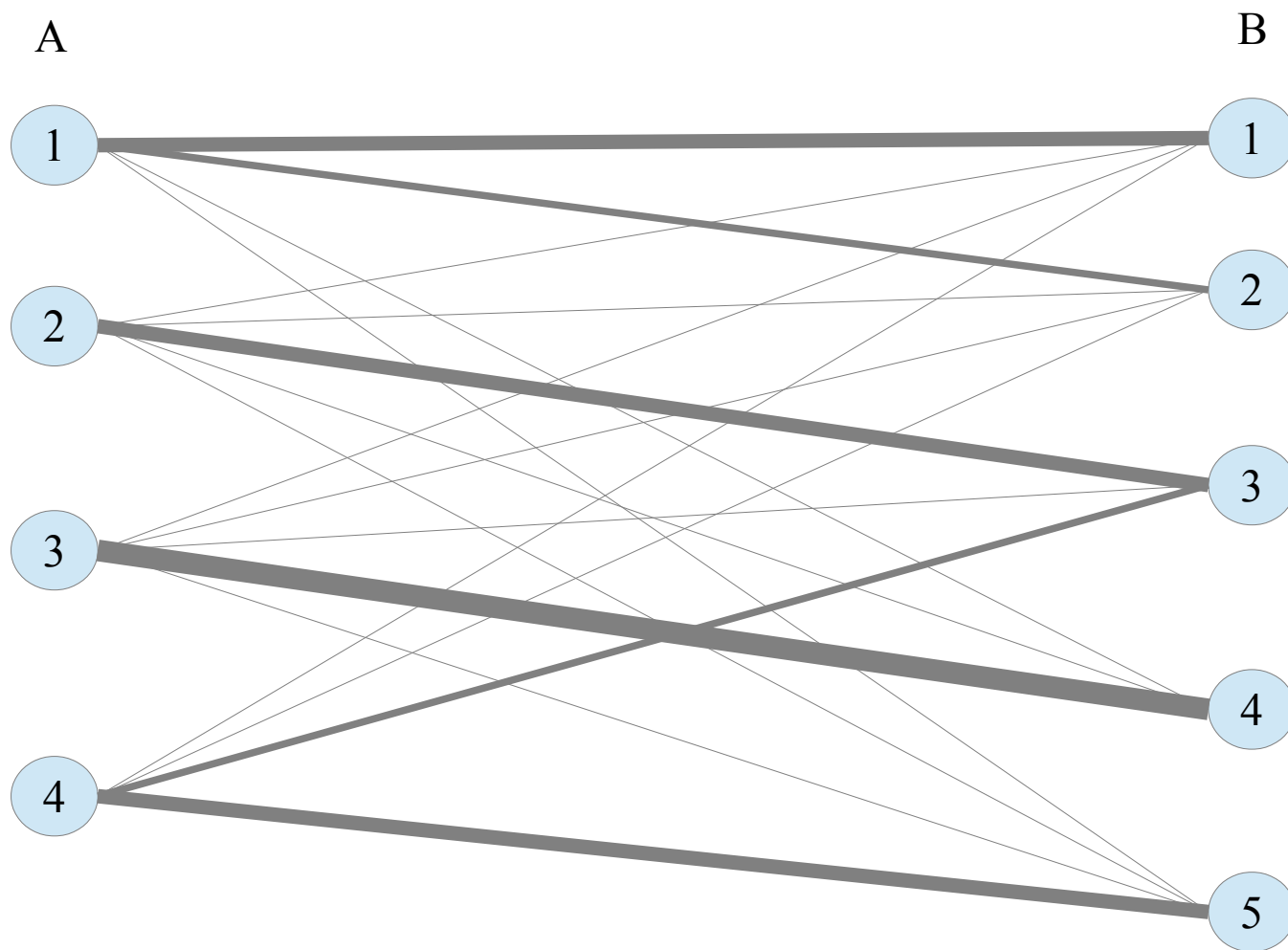


Simple superposition algorithm

```
Input: Structure A=x(1)..x(M); Structure B=y(1)..y(N)
Output: Best alignment Ali
Ali := {}      --- 初期アラインメント(空集合)
for (i,j,k) in {1..M} do --- Select 3 points from A
    basisA := make_basis(x(i),x(j),x(k)) --- Make a basis
    for a = 1..M do
        x'(a) := transform(x(a),basisA)
    for (l,m,n) in {1..N} do --- Select 3 points from B
        basisB := make_basis(y(l),y(m),y(n)) --- Make a basis
    S := {}      --- Initial (empty) alignment
    for b = 1..N do
        y'(b) := transform(y(b),basisB)

    (* After transformation, count neighboring A,B points *)
    for a = 1..M do
        for b = 1..N do
            if |x'(a) - y'(b)| < delta
            then S := S ∪ {(a,b)}      --- Add pair to alignment
if |S| > |Ali| then Ali := S      --- Save the best one!
```

A possible result

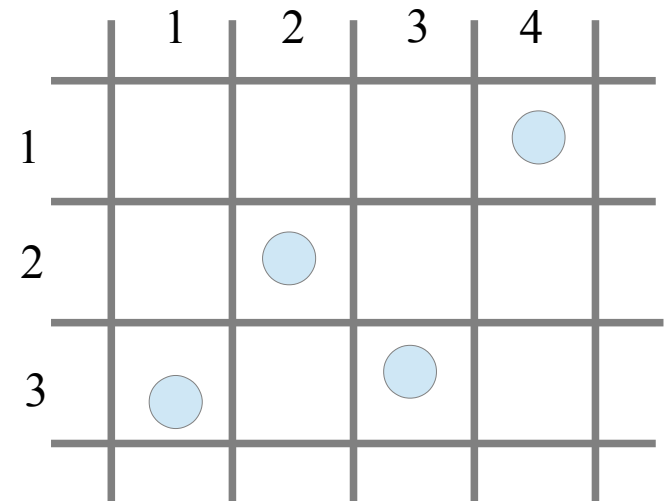


Geometric Hashing (GH)

- The simple approach is simply too slow.
- Make a dictionary (hash table) $x' \rightarrow \text{basis}$
- Looking up the dictionary is fast: $O(1)$, no loop.

The coordinate after transformed by basisB.

$$(x'(l), y'(l), z'(l)) \rightarrow \text{basisB}$$



Creating a hash table

Input: Structure B $y(1) \dots y(N)$

Output: Hash table HB

```
for (l,m,n) in 1..N do  
  basisB := make_basis(y(l),y(m),y(n))  
  for b = 1..N do  
    y'(b) := transform(y(b),basisB)  
    HB := HB      (y'(b) => y'(b),basisB)
```

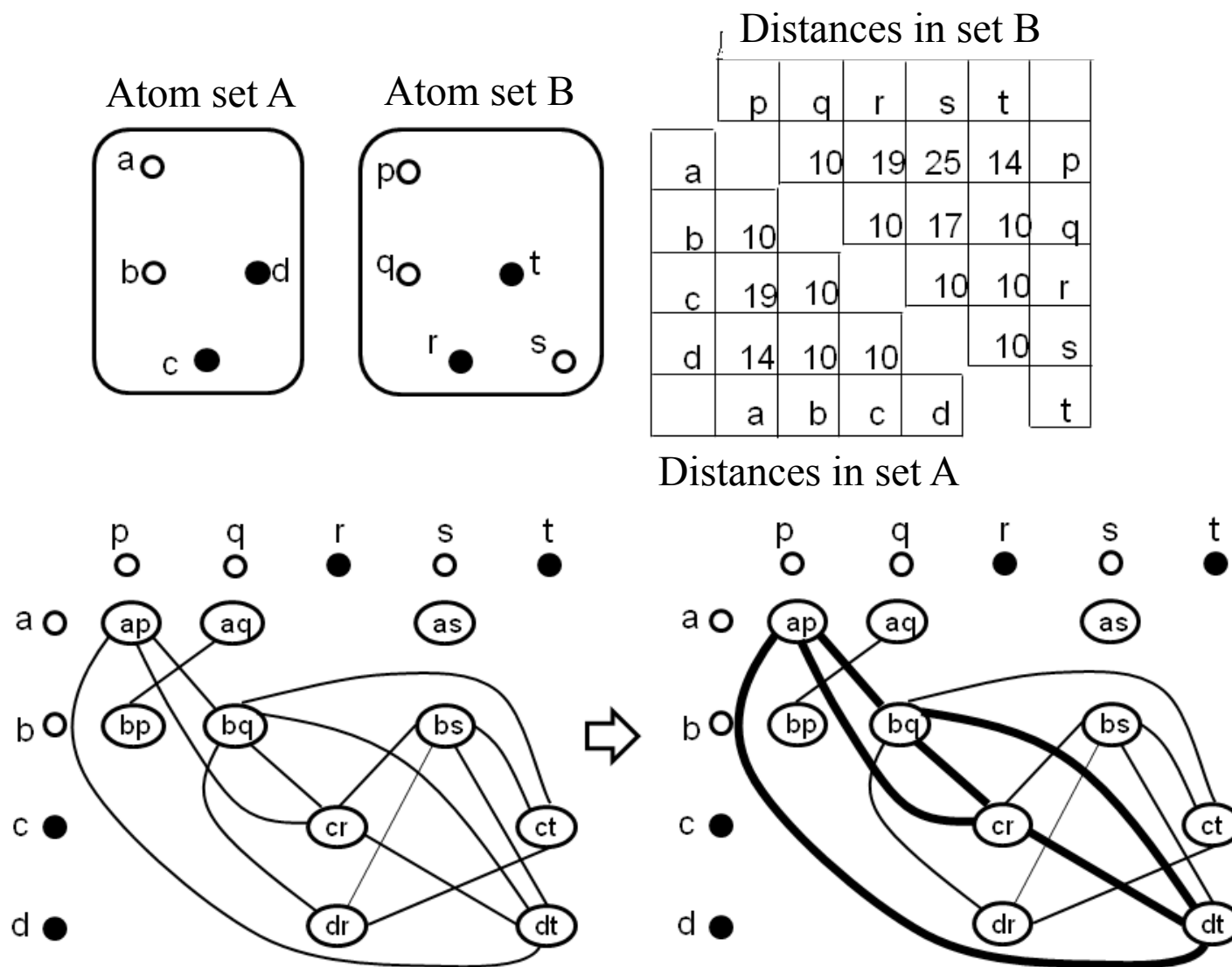
This requires $N^2(N-1)(N-2)$ steps.

Structure comparison by GH

```
Input: Structure A=x(1)..x(M); Structure B=y(1)..y(N)
Output: Best alignment Ali
HB := make_hashtable(B)  --- Create hash table
for (i,j,k) in {1..M} do --- Select 3 points from A
    basisA := make_basis(x(i),x(j),x(k))  --- Make a basis
    for a = 1..M do
        x'(a) := transform(x(a),basisA)
        for y'(b),basisB in find_hash(x'(a))  --- Find a B-basis
            P(basisA,basisB) := {(a,b)}      P(basisA,basisB)
            --- Add the atom pair
Ali := Max|P(basisA,basisB)|  ---(*) Be careful!
```

The last step (*) requires a smart data structure!
Otherwise, this method is as slow as the previous one.

Distance comparison method



Courtesy of Dr. Takeshi Kawabata

Basic idea of distance-based method

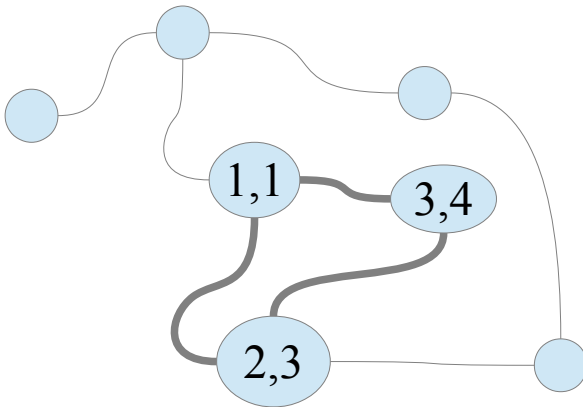
$A = (x_1^A, \dots, x_M^A)$
 $B = (x_1^B, \dots, x_N^B)$

Given A and B, consider all the pairs of A and B points: $P = \left\{ \left(x_i^A, x_j^B \right) \right\}$

For the pair of pairs (i,j) and (k,l), the two distances (i,k) & (j,l) are similar, draw an edge between the nodes (i,j) & (k,l).

$$\left| \left\| x_i^A - x_k^A \right\| - \left\| x_j^B - x_l^B \right\| \right| < \delta$$

Find the subgraph of thus created graph, that is complete and max imum:
 The maximum clique problem



Algorithm

Bron-Kerbosch (1973)

```
R := empty
P := set of vertices
X := empty
```

```
BronKerbosch1(R, P, X):
    if P and X are both empty:
        report R as a maximal clique
    for each vertex v in P:
        BronKerbosch1(R ∪ {v}, P ∩ N(v), X ∪ N(v))
        P := P \ {v}
        X := X ∪ {v}
```

Where $N(v)$ is the set of vertices connected with "v".

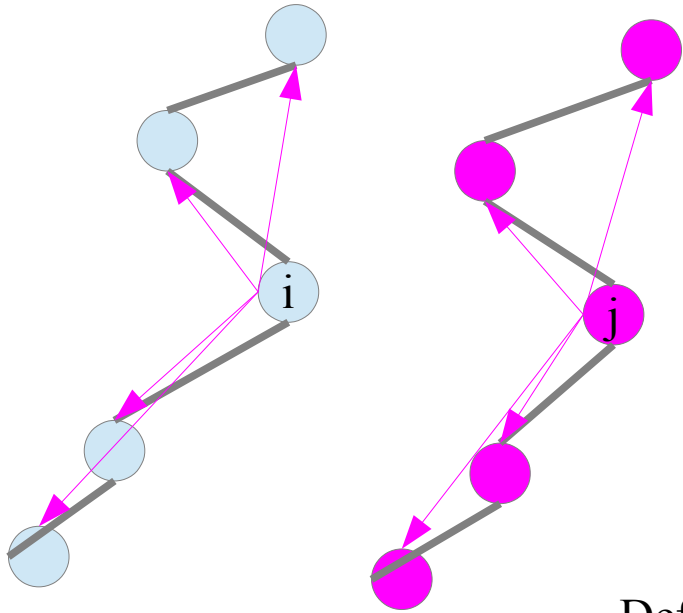
From http://en.wikipedia.org/wiki/Bron-Kerbosch_algorithm

This is an exact algorithm, and may not terminate.

Double Dynamic Programming

- Distance-based methods are also computationally demanding.
- DDP is a hybrid of coordinate- & distance-based methods
- Applying DP (just as in sequence comparison) in two layers.
- This requires the point set to be ordered.

DDP: idea



$$A = (x_1^A, \dots, x_M^A)$$

$$B = (x_1^B, \dots, x_N^B)$$

Assume (x_i^A, x_j^B) is a matching pair of points.

If (i,j) is really a matching pair, the “scene of A from i” and the “scene of B from j” should look similar.

Define the similarity measure for the “scenes” based on (i,j):

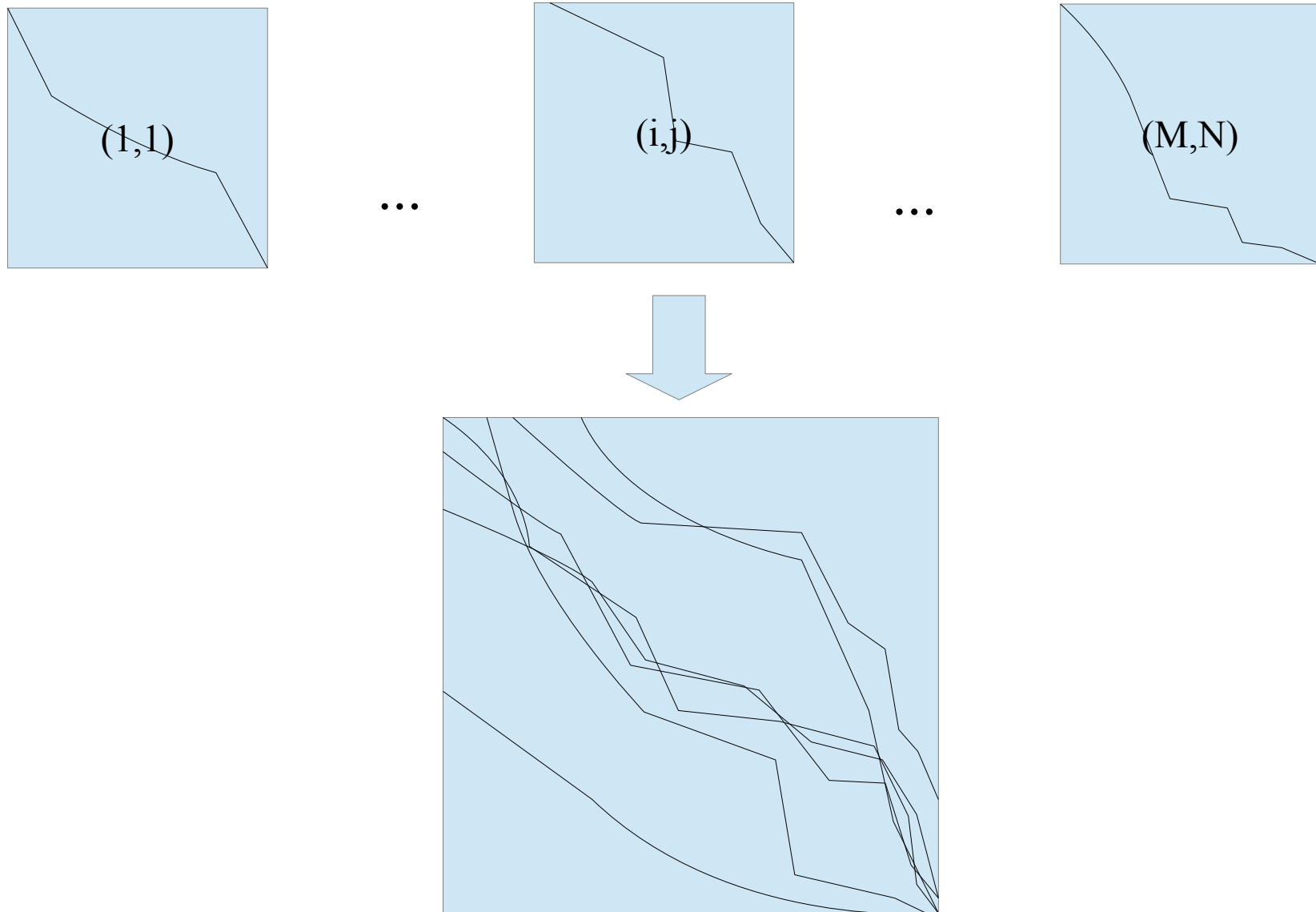
$$s(k, l; i, j) = \frac{1}{|d_A(i, k) - d_B(j, l)| + c}$$

Apply DP by regarding this as a score matrix $s(k, l)$, you get the “best” alignment under the assumption that (i,j) is a matching pair. The score is, say: $S_1(i, j)$ (Do this for all possible (i,j) pairs.)

Then using $S_1(i, j)$ as a score matrix, apply another DP.

This will yield an approximation to the “best” alignment

DDP の概念図



DDP Algorithm

```
# lower level DP
for i=1..M do
  for j=1..N do
    S(i,j) = DP using s(k,l; i,j)  --- details omitted.

# upper level DP
for i= 1..M do
  for j= 1..N do
    D := T(i-1,j-1) + S(i,j)
    V := T(i-1,j) - g
    H := T(i,j-1) - g
    T(i,j) := max(d,v,h)
    if T(i,j) = d then P(i,j) := 'D'          --- diagonal
    else if T(i,j) = v then P(i,j) := 'V'      --- vertical
    else T(i,j) := 'H'                        --- horizontal
  done
done
Score := T(M,N)
--- omitting the rest...
```

Why DDP works

- If (i,j) is a truly matching pair
 - $S_1(i,j)$ is a large positive value.
- If (i,j) is not a truly matching pair
 - $S_1(i,j)$ is a small value.
- The scores of truly matching pairs are amplified along the (sub)optimal alignment.

Summary

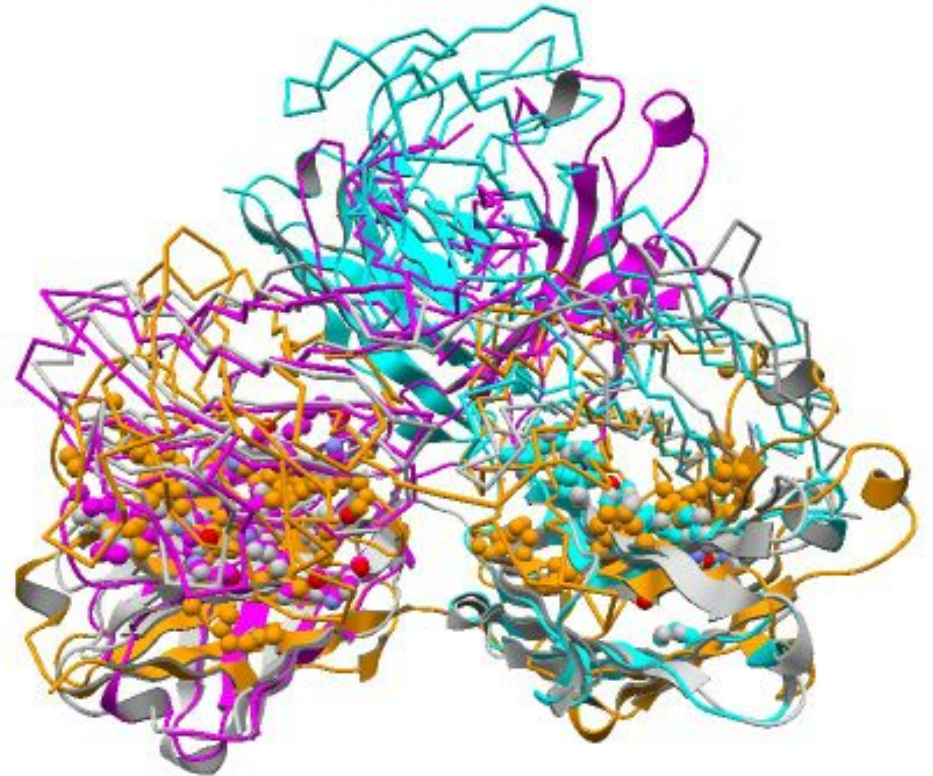
- 2 approaches for structure comparison
 - Coordinate-based
 - Distance-based
- In special cases, dynamic programming can be also used.

Application

Fast search and flexible alignment of
interaction site structures in proteins

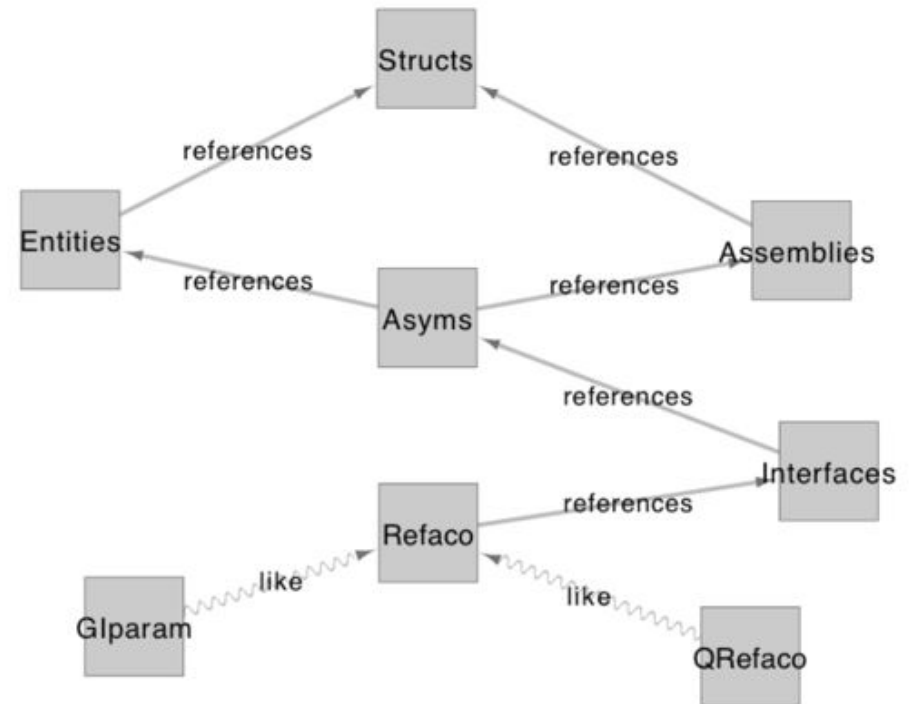
Objectives

- Search a large database of interaction site structures *quickly*.
- Align flexible structures



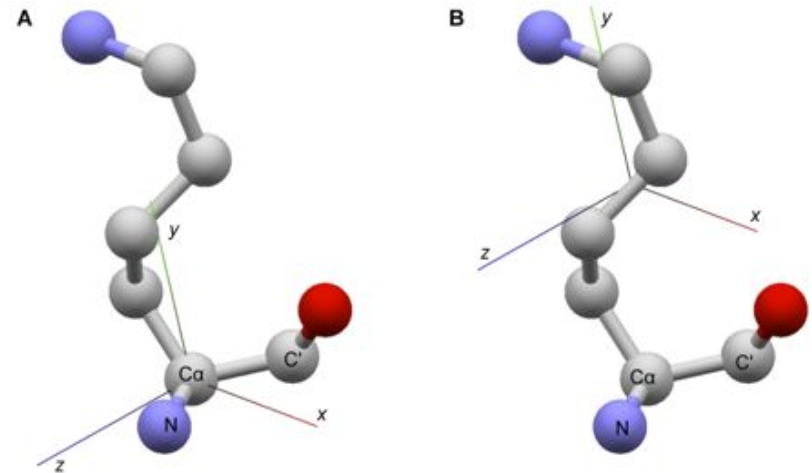
Database design

- From PDBML files, extract all interaction interfaces (small molecules, proteins, DNA/RNA)
- Biological assemblies are considered.
- Preprocess the local coordinates.



Local coordinate system

- origin: center of mass of side chain heavy atoms.
- axes: based on backbone N, C α , C' atoms.
- Defined for each residue in an interface.

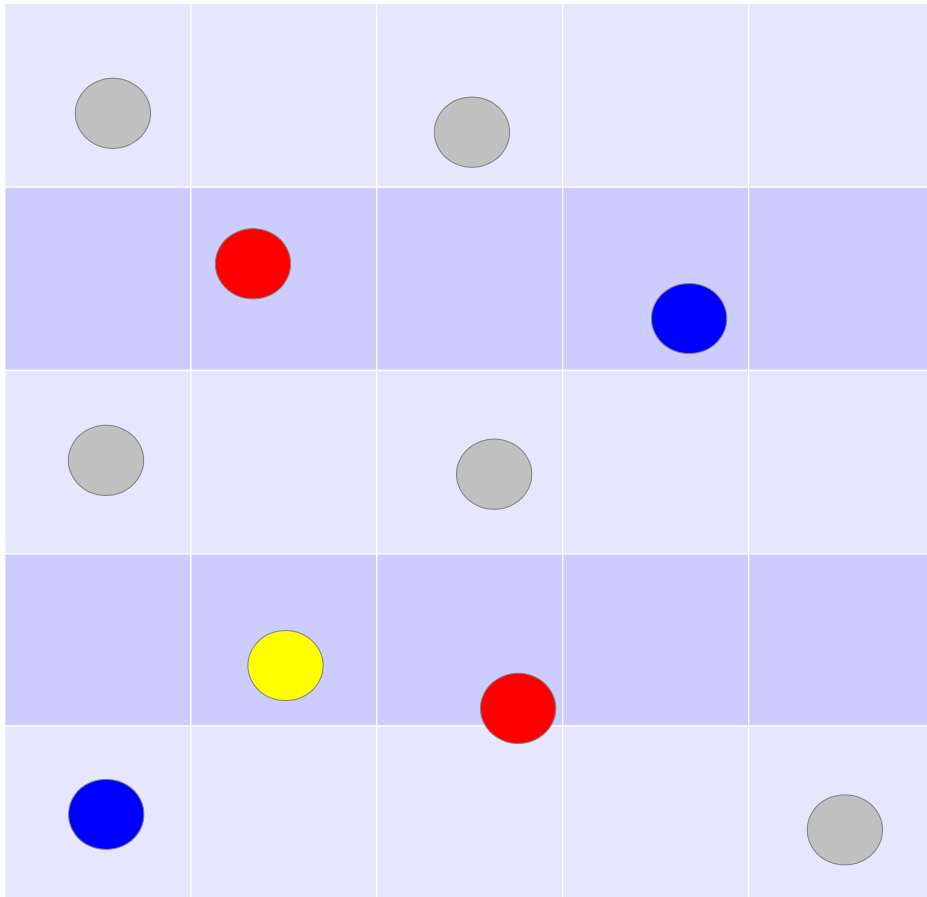


Structure search by SQL

```
CREATE TABLE Refaco (
  if_id TEXT          /* interface ID */
, rs_id TEXT          /* affine frame ID */
, type TEXT           /* interface type */
, frame BYTEA         /* affine frame */
, ft01 FLOAT          /* structural features 1~44 */
, ft02 FLOAT
, . . . . .
, ft44 FLOAT
, lattice BIGINT[ ] /* array of lattice points */
, natoms INT          /* number of interface atoms */
);
```

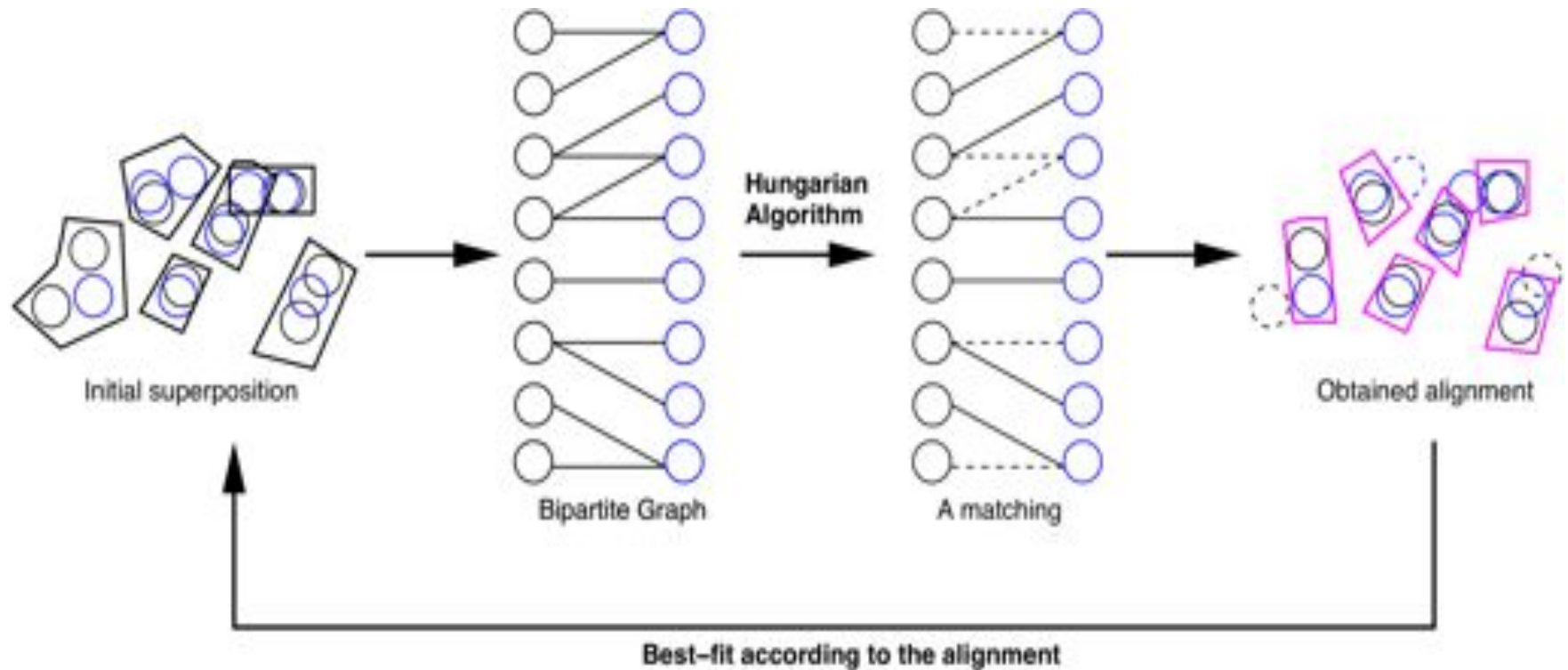
```
1: SELECT t.if_id, t.rs_id, t.type, t.frame, q.rs_id
2: FROM   Refaco t, Qrefaco q
3: WHERE  t.ft01 BETWEEN q.ft01 - D01 AND q.ft01 + D01
4: AND    t.ft02 BETWEEN q.ft02 - D02 AND q.ft02 + D02
        . . .
5: AND    t.ft44 BETWEEN s.ft44 - D44 AND q.ft44 + D44
6: AND    (SELECT COUNT(*)
7:         FROM (SELECT UNNEST(t.lattice)
8:               INTERSECT
9:               SELECT UNNEST(q.lattice)) AS x)
10:       > Smin * LEAST(t.natoms, q.natoms)
```

Discretized local atomic coordinates



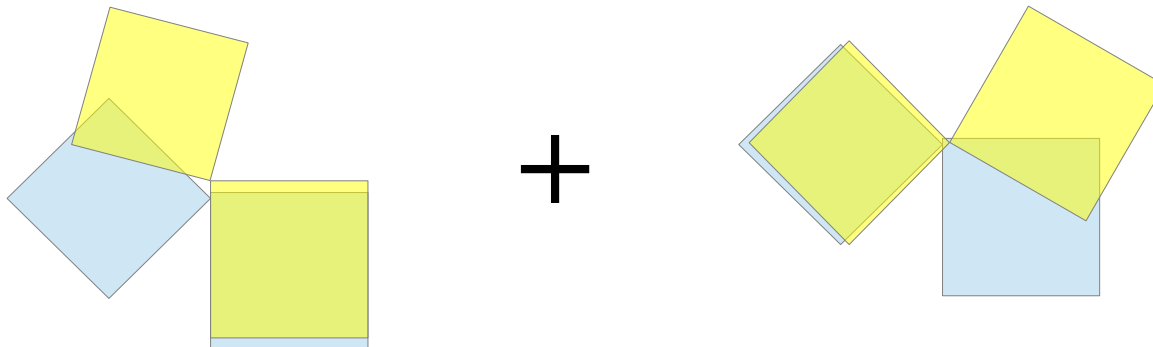
- Local coordinates of interface atoms are discretized and saved in an array-type column of the Refaco table.

Alignment by Hungarian method

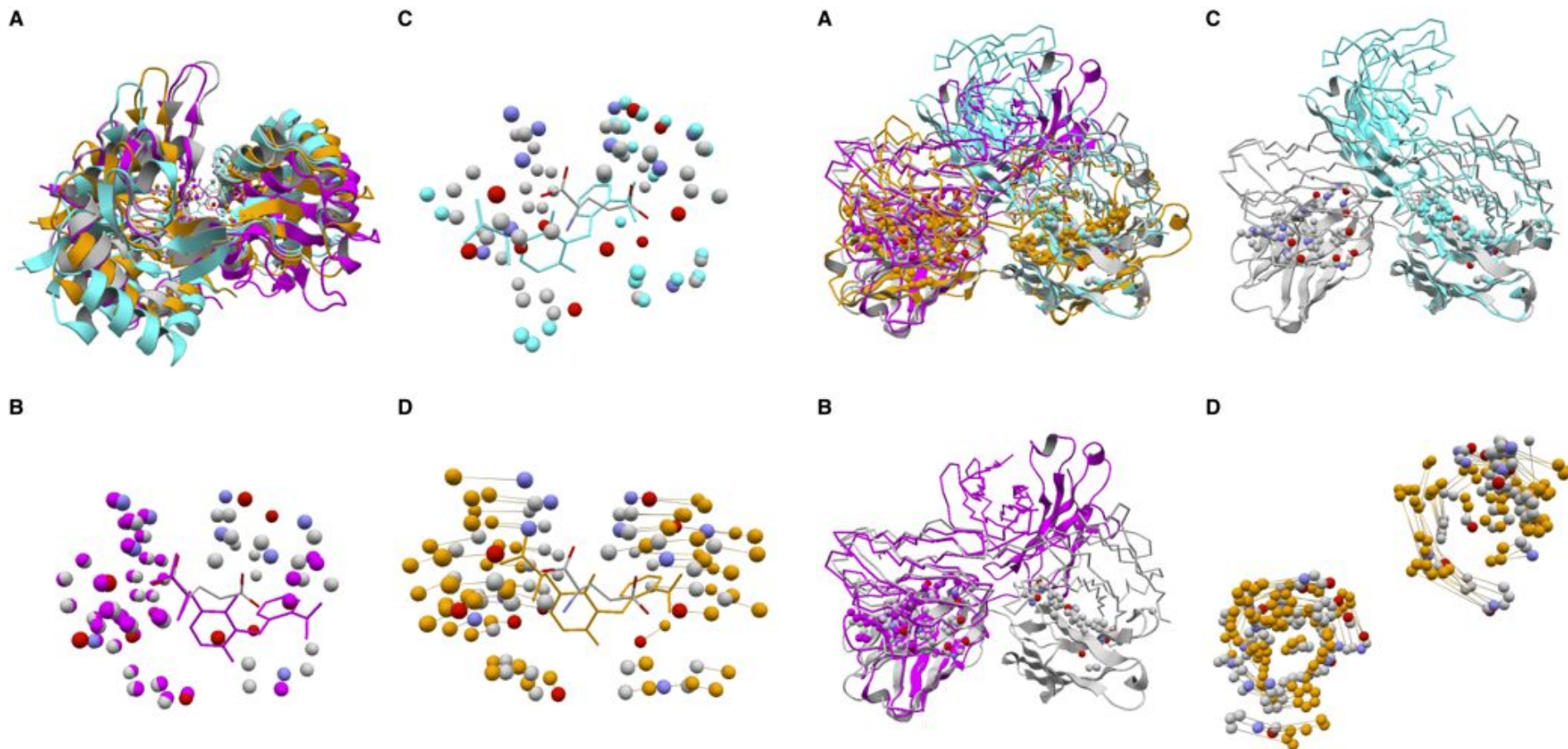


Flexible alignment

- Rigid alignments based on different affine frames.
- They are merged as long as mutually consistent.
 - Bad in 3D-Euclidean space, Good in patches of locally Euclidean spaces.



Flexible alignment: examples



Applications

Structure 17:234-246 (2009)

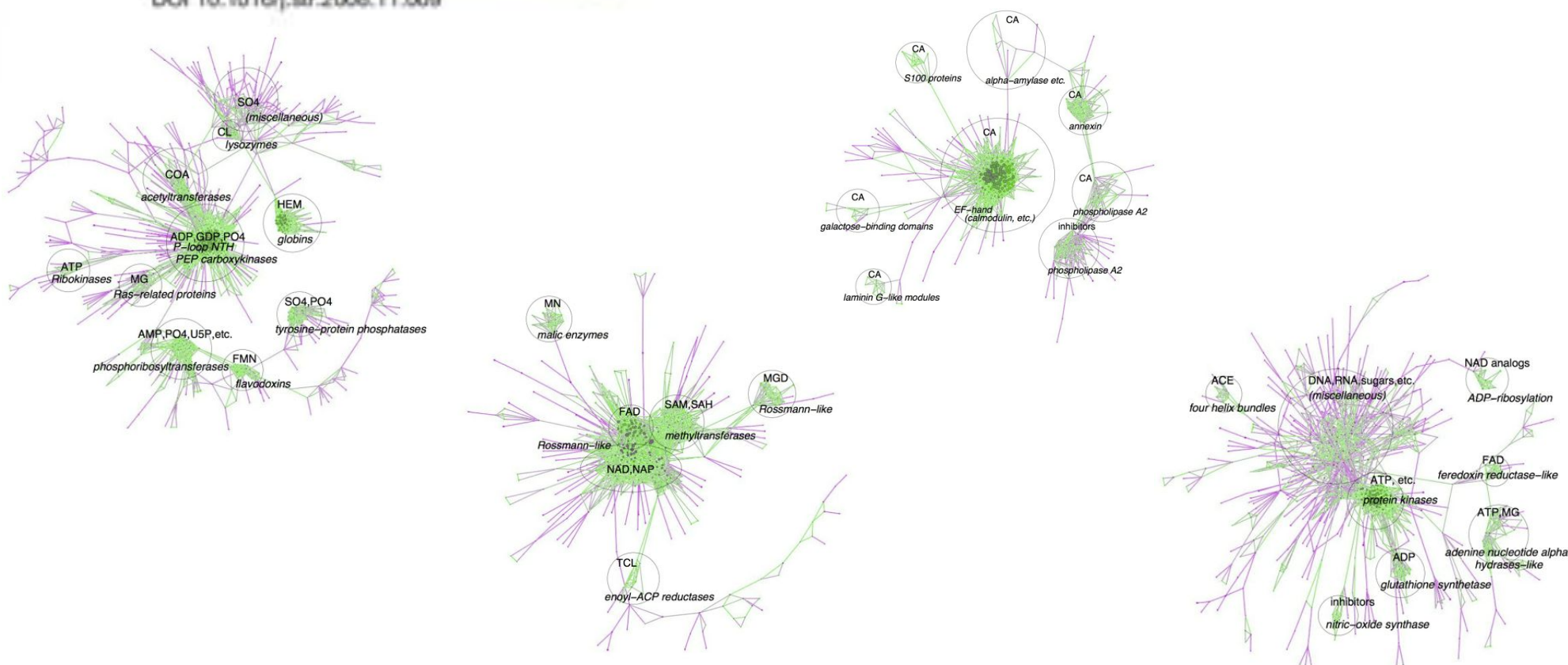
Comprehensive Structural Classification of Ligand-Binding Motifs in Proteins

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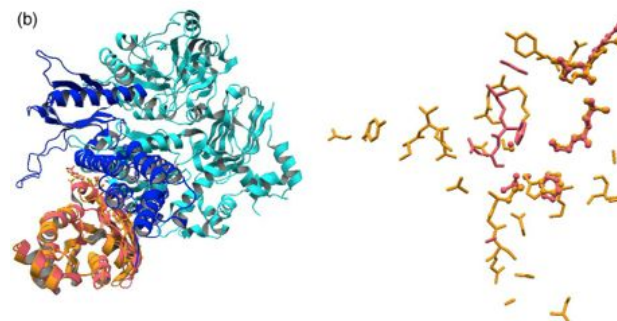
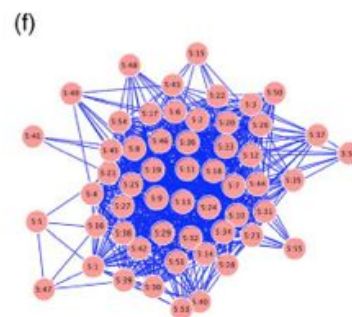
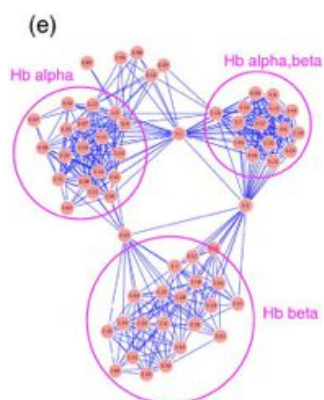
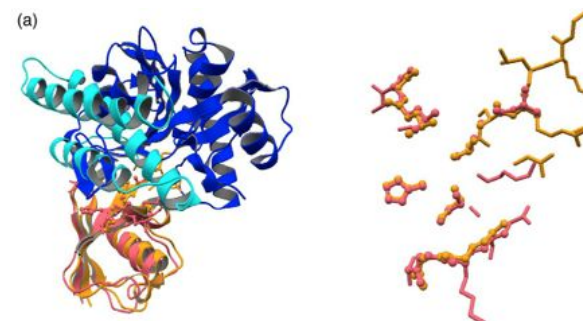
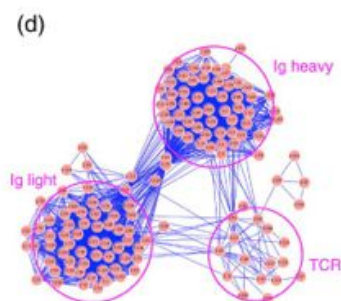
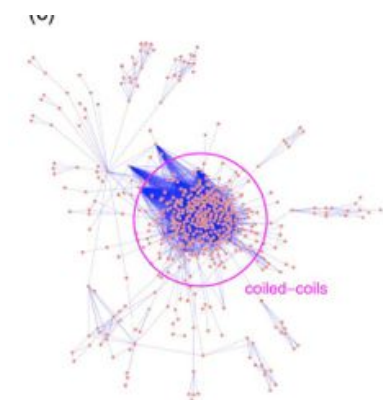
DOI 10.1016/j.str.2008.11.009





Geometric Similarities of Protein–Protein Interfaces at Atomic Resolution Are Only Observed within Homologous Families: An Exhaustive Structural Classification Study

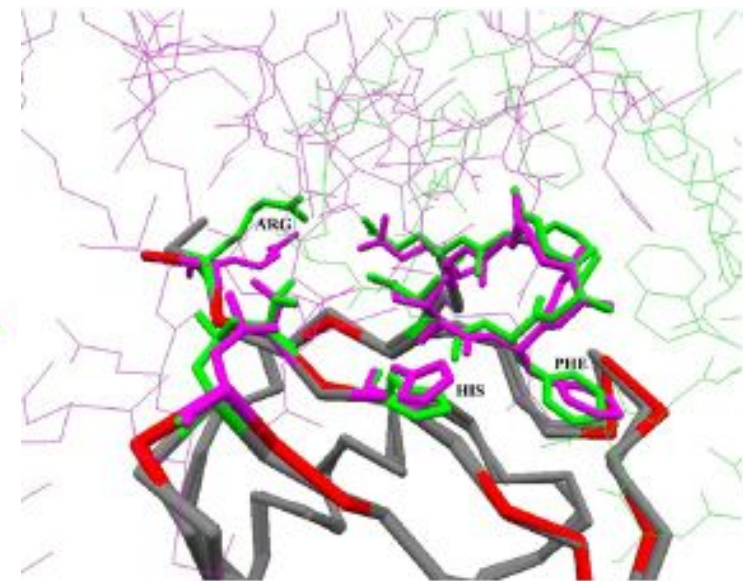
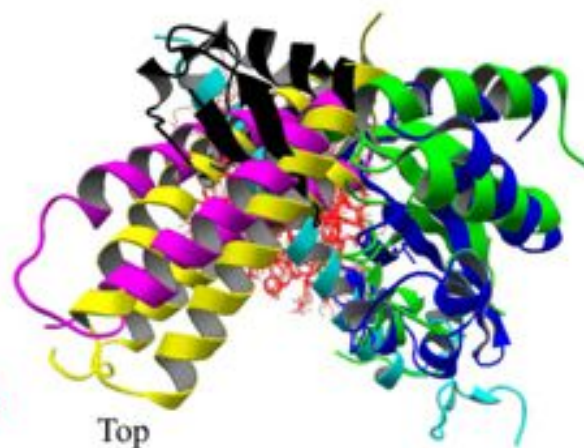
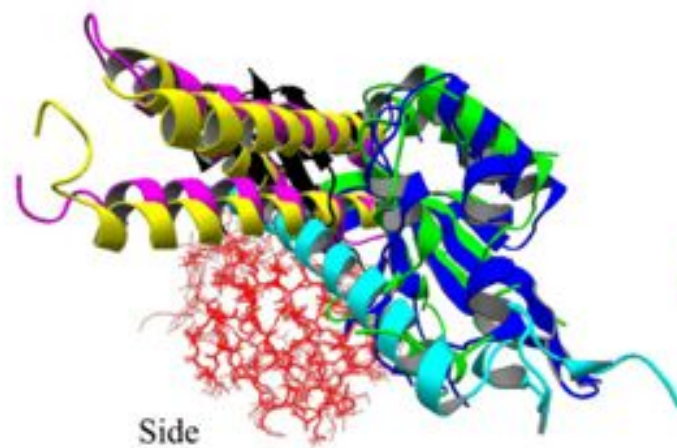
Akira R. Kinjo* and Haruki Nakamura





Distinct Roles of Overlapping and Non-overlapping Regions of Hub Protein Interfaces in Recognition of Multiple Partners

Bhaskar Dasgupta^{1,2*}, Haruki Nakamura¹ and Akira R. Kinjo¹



Composite Structural Motifs of Binding Sites for Delineating Biological Functions of Proteins

Akira R. Kinjo*, Haruki Nakamura

PLoS ONE 7(2): e31437 (2012)

What is “protein function”?

- Biochemical function
 - Catalytic activity (enzymes), ligand binding...
 - Attributes of proteins themselves.
- Biological function
 - “Development”, “aging”, “memory”,...
 - Behavior of a network of proteins.

How do you know a function?

- Adenylate kinase 1 (human) in UniProt:

General annotation (Comments)		Hide Top
Function	Catalyzes the reversible transfer of the terminal phosphate group between ATP and AMP. Small ubiquitous enzyme involved in energy metabolism and nucleotide synthesis that is essential for maintenance and cell growth.	
Catalytic activity	$\text{ATP} + \text{AMP} = 2 \text{ ADP}$.	
Subunit structure	Monomer.	
Subcellular location	Cytoplasm.	
Polymorphism	This enzyme represents the most common of at least five alleles.	
Involvement in disease	Defects in AK1 are the cause of hemolytic anemia due to adenylate kinase deficiency [MIM:612631].	
Sequence similarities	Belongs to the adenylate kinase family.	

Another way to annotate adenylate kinase in *Gene Ontology*

GO:0004017 adenylate kinase activity

Catalysis of the reaction: ATP + AMP = 2 ADP.

[Term Information](#) [Ancestor Chart](#) [Ancestor Table](#) [Child](#)

[Terms](#) [Protein Annotation](#) [Co-occurring Terms](#)

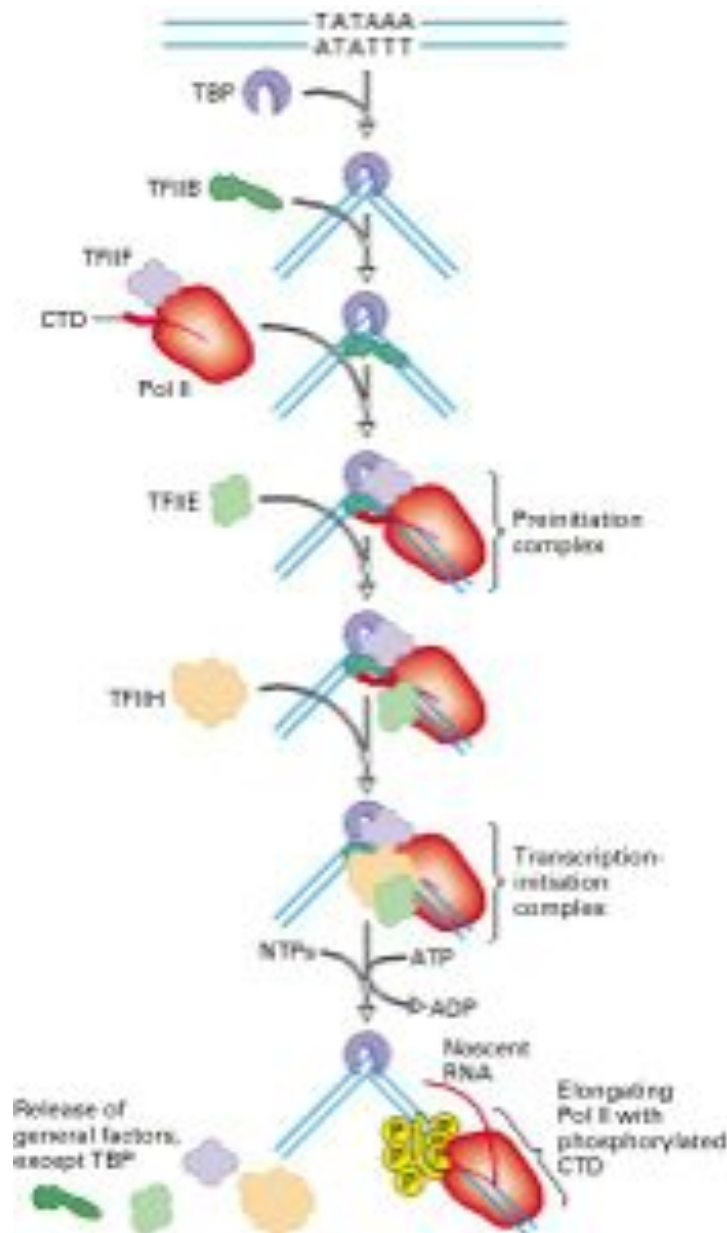
ID	 GO:0004017
Name	adenylate kinase activity
Ontology	Molecular Function
Definition	Catalysis of the reaction: ATP + AMP = 2 ADP.
Comment	
Secondary IDs	

[GONUTS Wiki Page](#)

[Synonyms](#)

Type	Synonym
exact	ATP:AMP phosphotransferase activity
narrow	myokinase activity
exact	adenylokinase activity
exact	adenylic kinase activity
exact	5'-AMP-kinase activity

What is protein function?



From *Molecular Cell Biology* 4/e (Fig. 10-50)

Protein function is ...

**A series of
Interaction States,
which is ...**

An ordered set of
binding patterns

Connecting
atomic structures
to
*higher-order biological
functions*

Plan

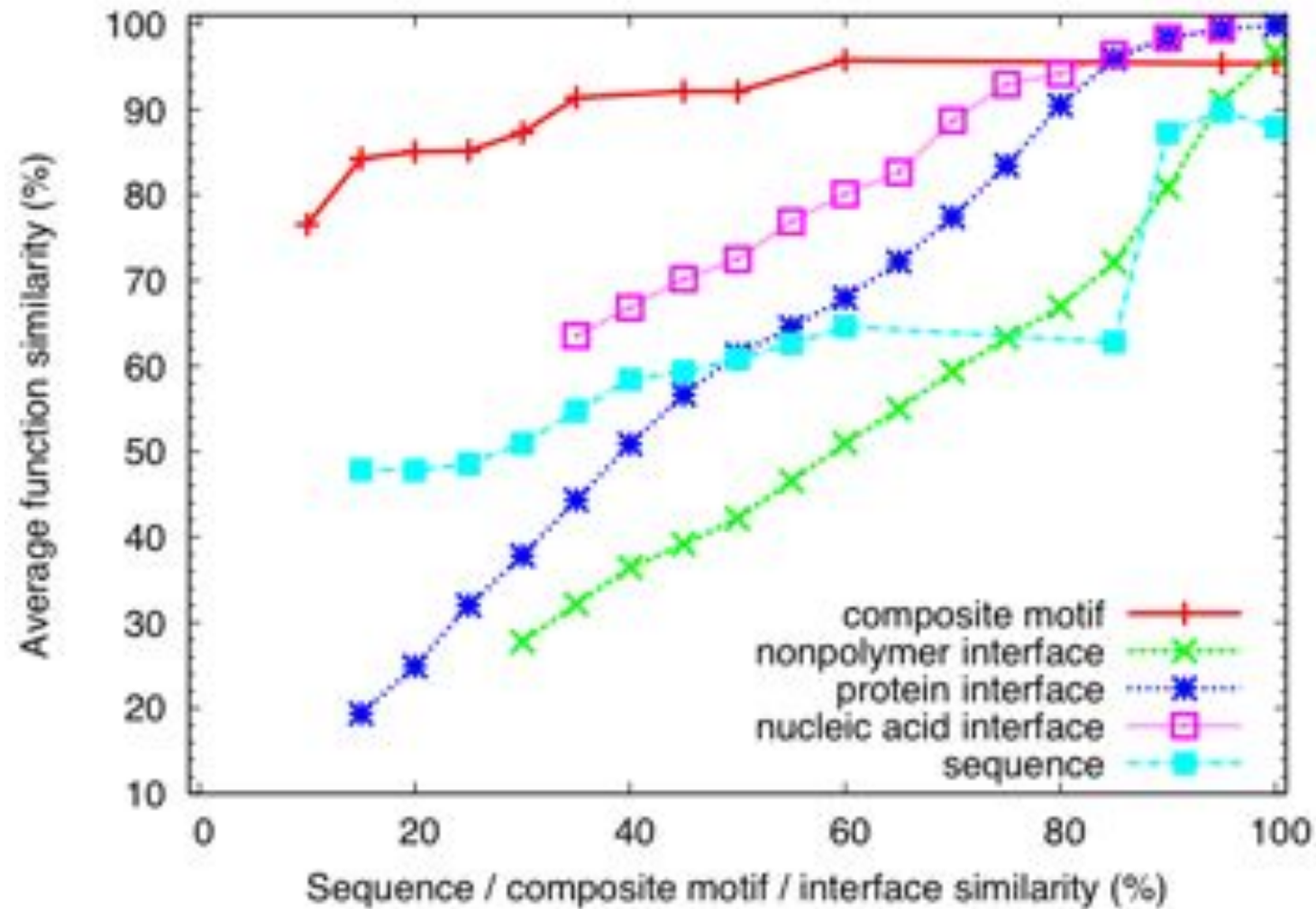
- Identify all patterns of binding sites
 - elementary motifs
- Identify all combinations of elementary motifs in each subunit
 - composite motifs
- Identify all composite motifs associated with a particular protein function (defined by UniProt).
 - meta-composite motifs
 - ~ a network of interaction patterns.

Materials

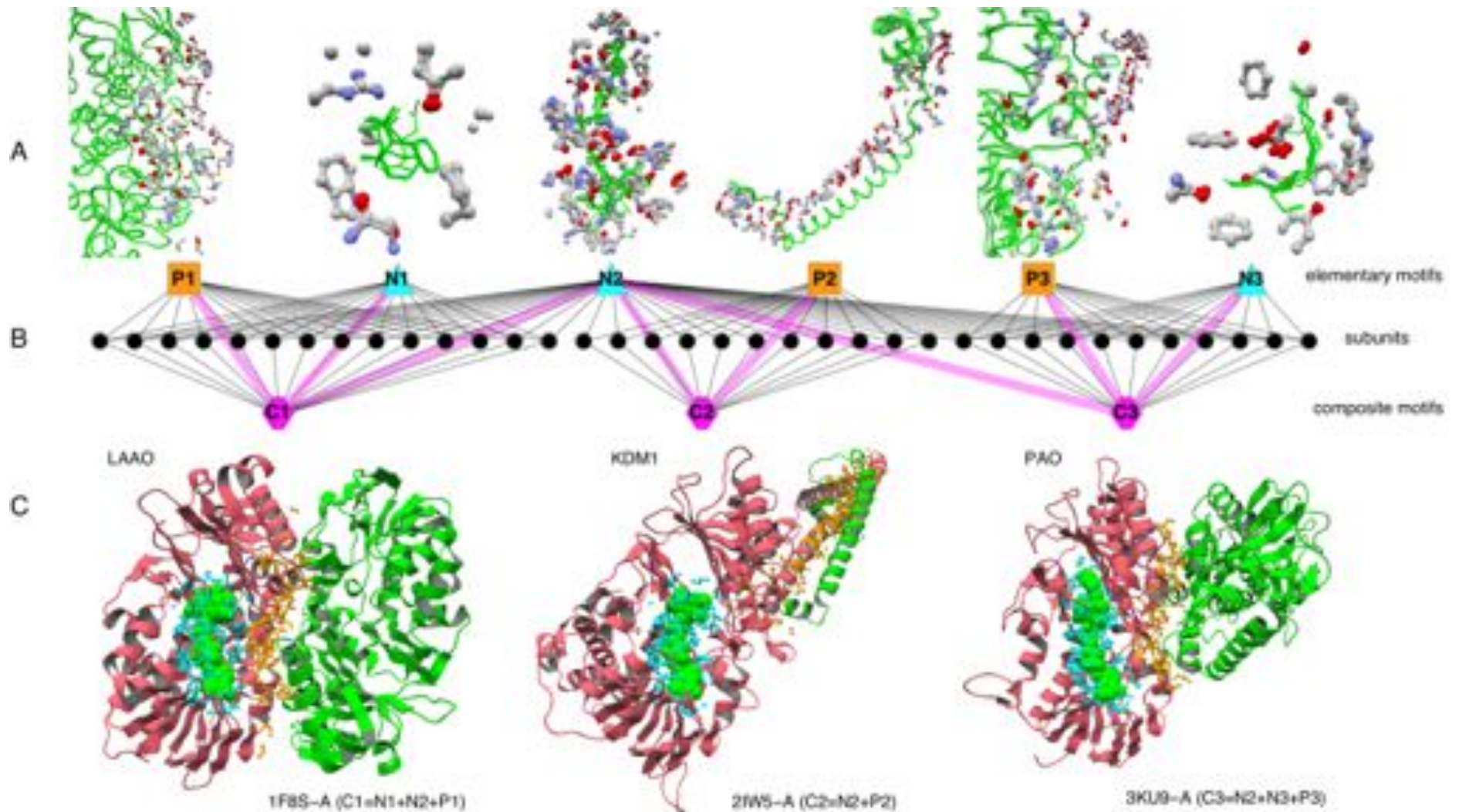
PDB entry	~70,000	
Biological Unit	~80,000	
Subunits	~200,000	~6,000 composite motifs
non-polymer interface	~400,000	~6,000 elementary motifs
protein interface	~350,000	~8,000 elementary motifs
DNA/RNA interface	~20,000	~400 elementary motifs

No sequence representatives are used!

c-motifs vs. functions



Composite motifs: example



Examples

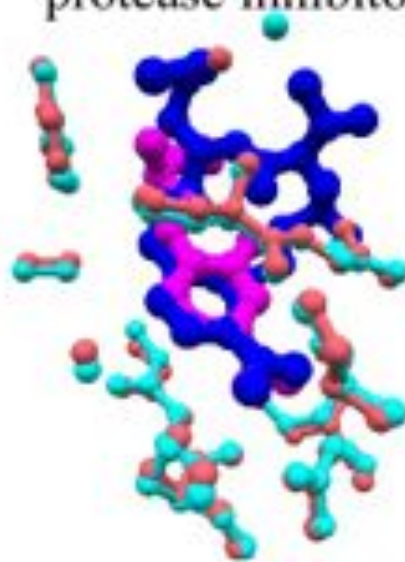
beta-trypsin (EC 3.4.21.4)



Factor VII (EC 3.4.21.21)



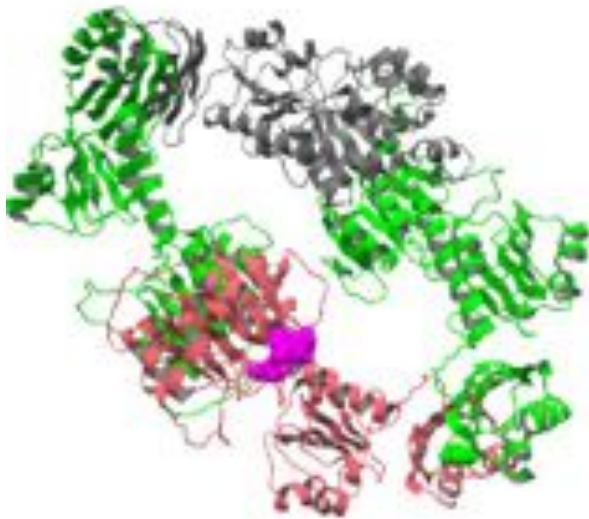
protease inhibitor



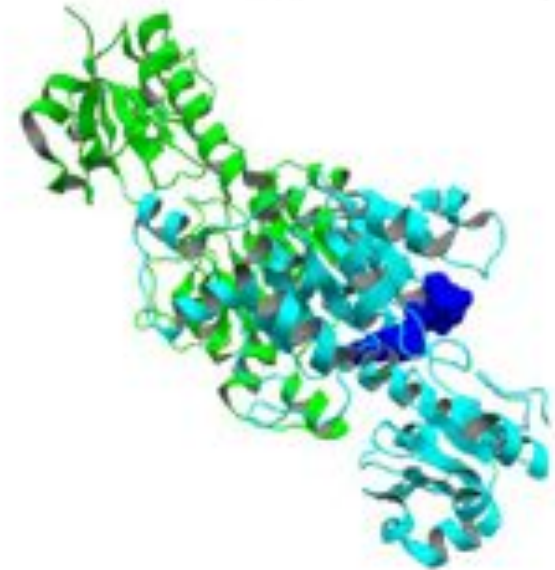
(i.e. catalytic site)

Same e-motif & fold, different c-motifs & functions

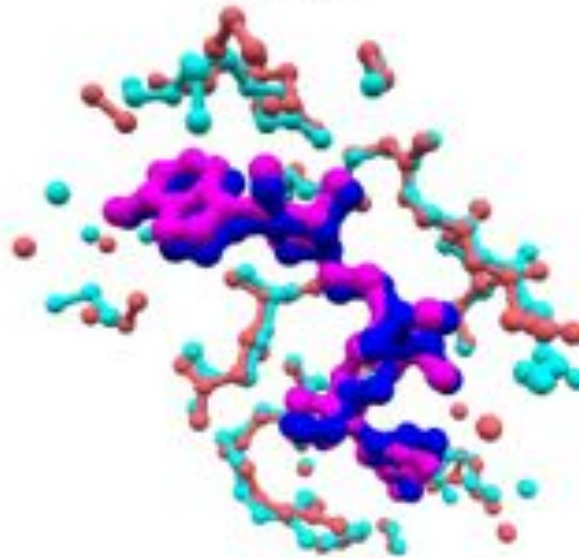
PGDH (EC 1.1.1.95)



CtBP3 (EC 1.1.1.—)

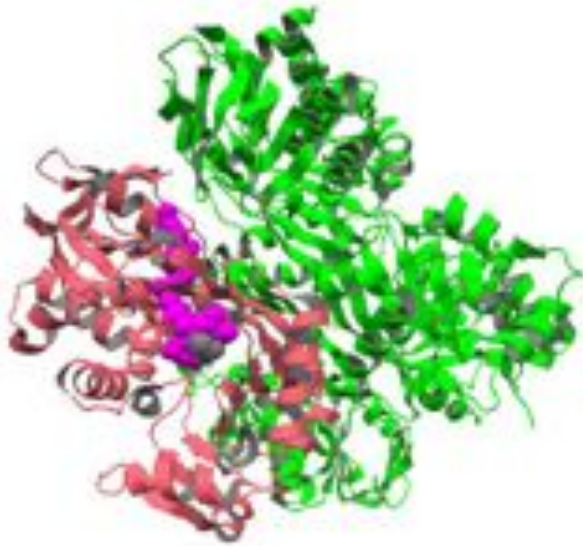


NAD

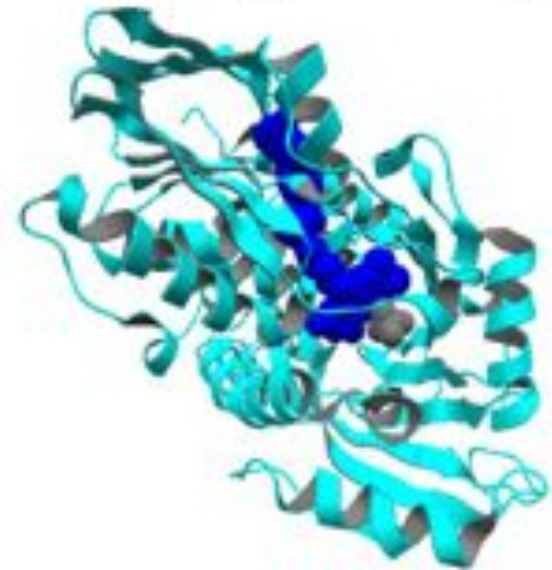


Same e-motif & fold, different c-motifs & functions

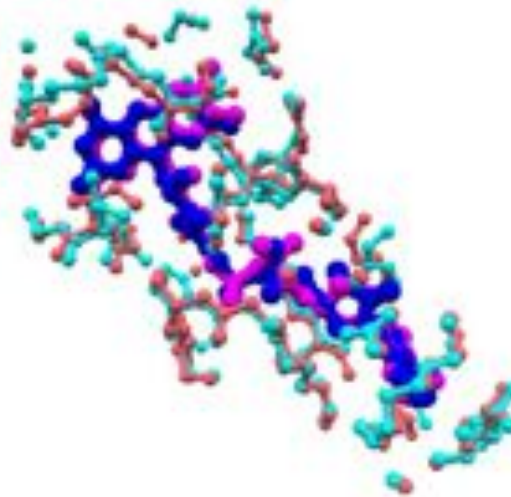
GO (EC 1.4.3.19)



GlpD (EC 1.1.5.3)

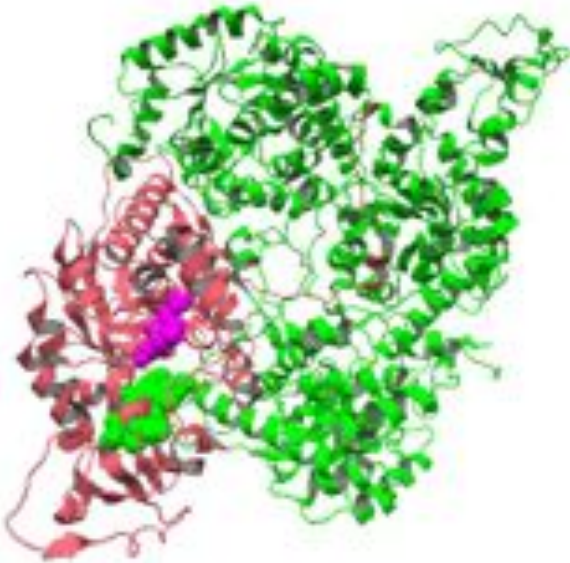


FAD

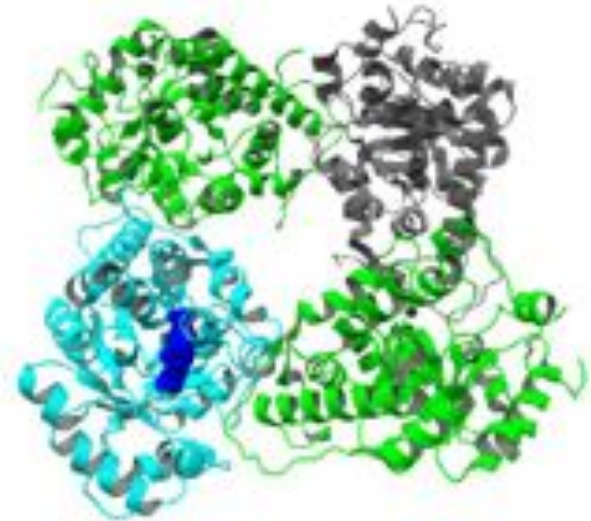


Same e-motif & fold, different c-motifs & functions

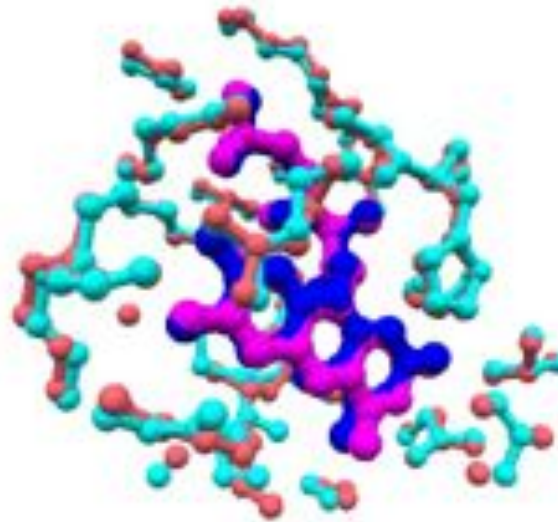
Cyt. b2 (EC 1.1.2.3)



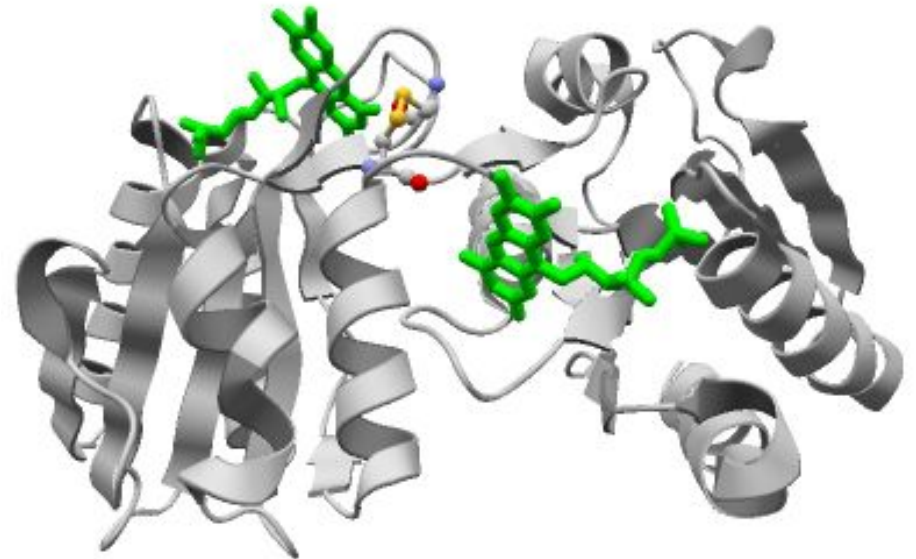
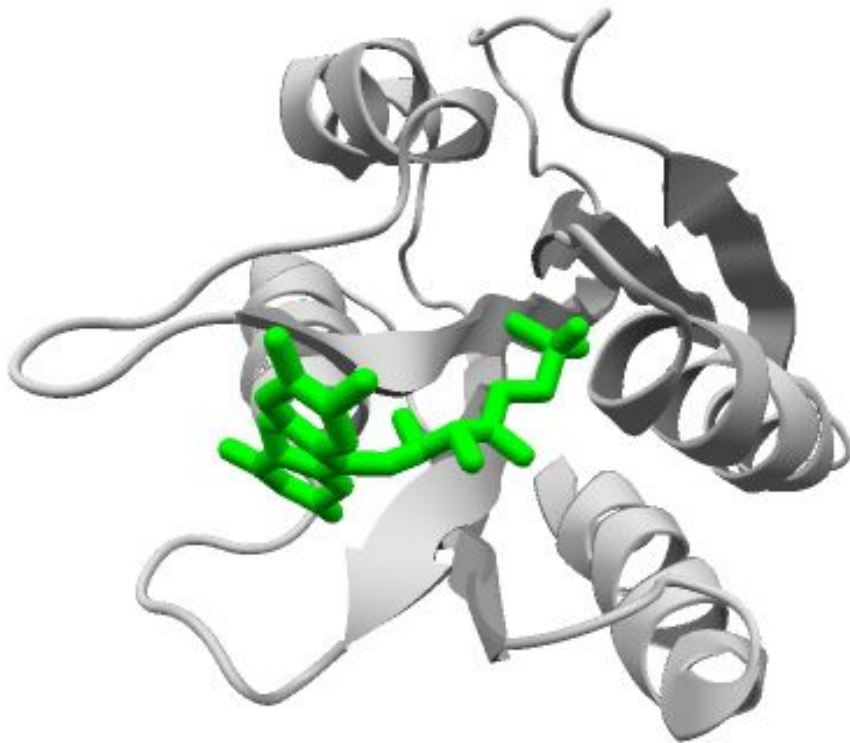
GOX (EC 1.1.3.15)



FMN



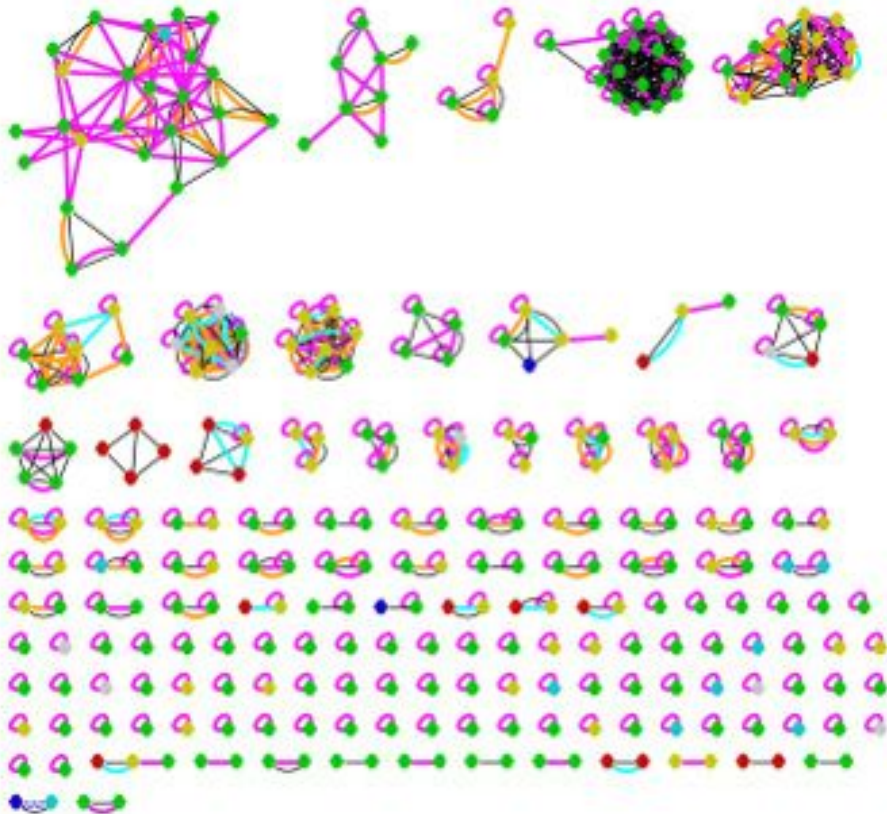
Flavodoxin: monomer & dimer



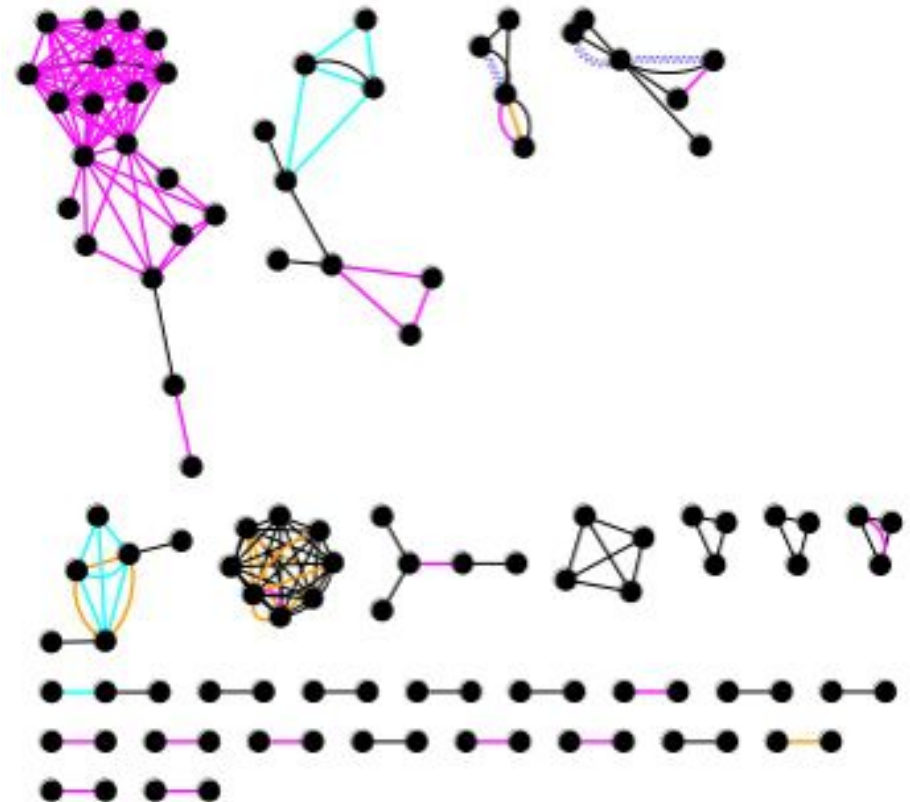
Meta-motif networks

“Transcription”

Meta-composite motif

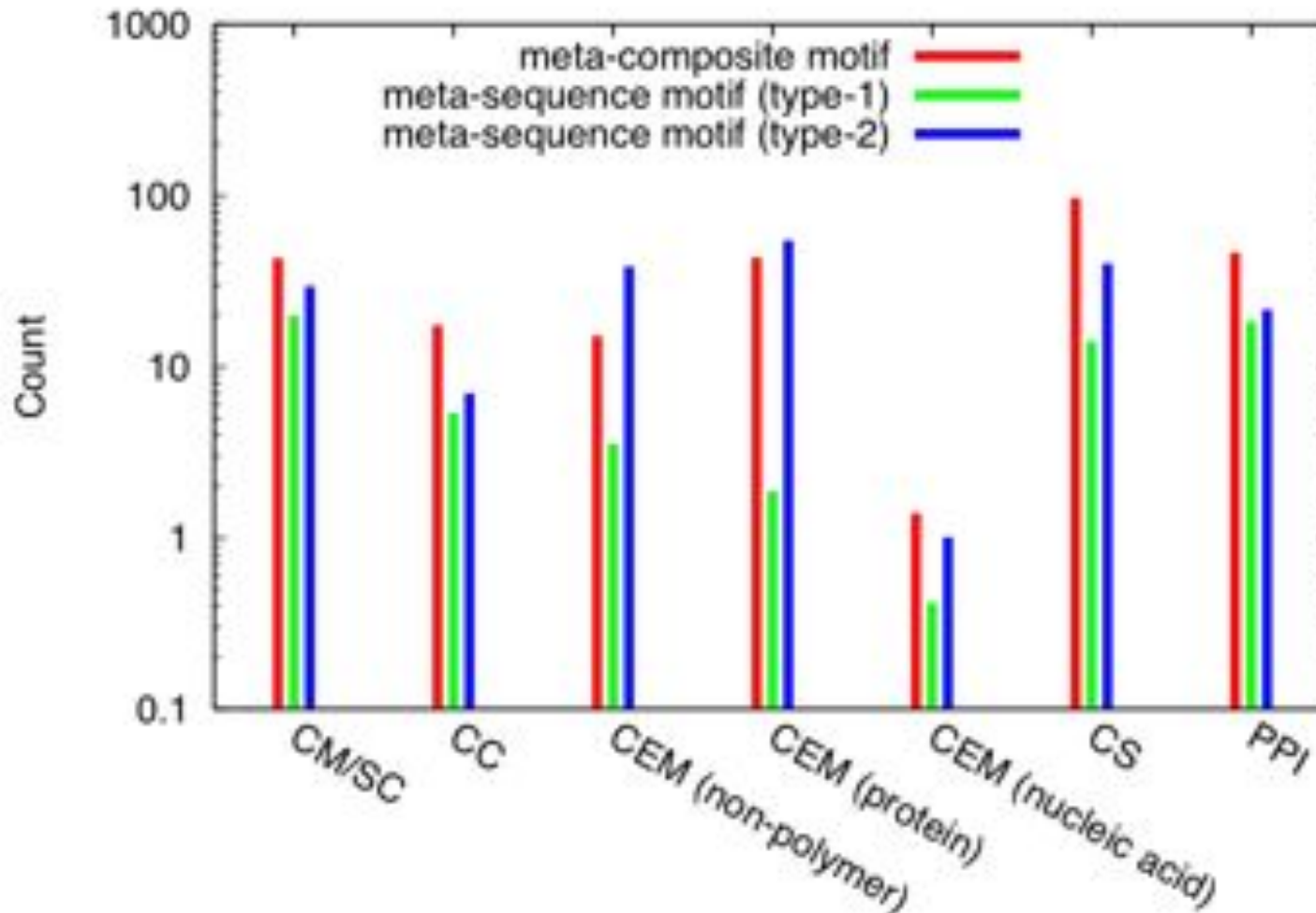


Meta-sequence motif



- node colors indicate interaction states of c-motifs.
- edges colors indicate relations between nodes.

Network characteristics



type-1 sequence clusters include remote homologs (BLAST e-value < 0.01)
type-2 sequence clusters consists only of close homologs (100% SID)

Conclusion

- Composite motifs better distinguish functions
 - different c-motifs $\Rightarrow$ different functions
 - “Difference” rather than “Similarity”
- Meta-composite motifs provide richer annotations of biological processes.
 - Transitions between interaction/conformational states
 - But time-dependence is still ignored...